

Original Research Article

Levels of Interleukin-6,17, Complement and Dopamine in Infected Women with Toxoplasmosis

Talib Wseen Hussein¹, Khalil Ismail Abid Mohammed^{2*}, Mohammed Abdul Salam¹

¹Department of Sciences/College of Basic Education/University of Al -Mustansyria/Baghdad-Iraq

²Clinical Communicable Diseases Research Unit /College of Medicine /University of Baghdad/Baghdad-Iraq

*Corresponding Author: Khalil Ismail Abid Mohammed

Clinical Communicable Diseases Research Unit /College of Medicine /University of Baghdad/Baghdad-Iraq

Article History

Received: 25.04.2022

Accepted: 31.05.2022

Published: 05.06.2022

Abstract: The study was carried out to detection of *Toxoplasma gondii* in (135) women patients with repeated abortion with age ranged 18-43 years who attended two teaching hospitals in Baghdad. The diagnosis is done by Immunochromatography and ELISA methods. a blood samples was taken from each patients as well as other(30)healthy control matching in age. The study included measurement the concentration of Interleukin-6, Interleukin-17, Dopamine and Complement in sera of patients and control .The result indicated presence the of anti-Toxoplasma IgG in 26 cases a, Anti-toxoplasma IgM in 14 cases and 10 in both out of 135 cases of aborted women with Toxoplasma in immunochromatography methods while the level of IgM 1.57 IU/ml and IgG 142.73. Also, the result indicated significant increasing levels of IL-6 and IL-17, Complement and Dopamine in patients sera in comparison with healthy control.

Keywords: Toxoplasmosis, Interleukin -6, Interleukin -17, Dopamine.

INTRODUCTION

Intracellular parasites like *Toxoplasma gondii*, usually contaminate with an extensive level of infections to the eukaryotic cells. It is quite significant devious pathogenic organism in both humans as well as animals. Often this contamination comes with no symptoms, nevertheless, the two sets that threaten personally, the humanoid genes and the persons with immunosuppression, mainly to those who have weak immune system, which lead to lethal toxoplasmic encephalitis [1]. Parasites duplicate gradually and eventually form bradyzoite that remain in the cysts of the tissues of the neural hosts as well as the tissues of the muscles for the host life time [2].

The inherited contamination could lead to defects in birth which includes hydrocephalus, spontaneous abortion, chorioretinitis and intra cerebral calcification [3]. The innate immune system responds through monocytes, dendritic cells (DCs) and T. *gondii* which ends up infecting tissues. These types of cells are involved and are responsible for the microbial resistance [4-9] *T. gondii* is the ability to sense the pathogen and produce the cytokine IL-12, which stimulates natural killer (NK) cells and T cells to produce the cytokine Interferon-gamma (IFN- γ) [10–12]. IFN- γ is the major mediator of resistance to *T. gondii* and promotes multiple intracellular mechanisms to kill the parasite and inhibit its replication. This Th1 immune response, defined by the production of IL-12 and IFN- γ , is characteristic of infection with many intracellular pathogens [12, 13].

Neutrophils are another source of IL-12 during toxoplasmosis, as they contain pre-stored IL-12 and can secrete this cytokine in vitro and in vivo in response to *T. gondii* [14]. Additionally, there are reports that neutrophil depletion results in decreased levels of IL-12 and increased parasite replication [15]. Natural killer (NK) cells are another innate population involved in immunity to *T. gondii*, and in mice that lack T cells they provide a limited mechanism of resistance through their ability to produce IFN- γ [16–18] NK cell activity peaks early during infection, and although their activity is elevated during chronic toxoplasmosis, they do not appear to be significant contributors to immunity during

Copyright © 2022 The Author(s): This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International License (CC BY-NC 4.0) which permits unrestricted use, distribution, and reproduction in any medium for non-commercial use provided the original author and source are credited.

CITATION: Talib Wseen Hussein, Khalil Ismail Abid Mohammed, Mohammed Abdul Salam (2022). Levels of Interleukin-6,17, Complement and Dopamine in Infected Women with Toxoplasmosis. *South Asian Res J Pharm Sci*, 4(3): 54-59.

the chronic stage of infection [19, 20]. NK cells can also act to promote adaptive immune responses. Thus, in the absence of CD4⁺ T cells, they can provide help to the CD8⁺ T cell response [21].

MATERIAL AND METHODS

Study groups

This research work was conducted in two teaching hospitals in Baghdad the patients' ages were between 18 to 43 years old. The patients were divided into two groups, which include suspected patients. Blood samples were collected from 135 women suspected as *T. gondii* infected which was tested and confirmed (clinically suspected cases) by specialist doctors and the other group was control including healthy individuals.

Blood Samples

The blood samples were taken from the participants' veins and were collected in pastoralized plastic tubes. Each sample was left for 30 minutes at room temperature in serum separator tubes in vertical position prior to centrifugation. The samples were centrifuged at 3000 rpm and kept for 5 min. The serum was collected from every sample using Eppendorf tube and was stored at -20°C. This sample was used to evaluate immunological and clinical biochemical aspects.

Immunochromatographic assay

About 100 µl of serum from each sample was added to the sample hole of the kit. The colour density is proportional to the antibody titer. The complexes (appears in colour band after 10 minutes) confirm that the test was performed correctly. This CerTest-Toxoplasma kit which is qualitatively determines the Toxoplasma in blood samples. Pre-coating was achieved to the membrane proceeding to test band region to the monoclonal antibodies of the mouse, it was achieved against Toxoplasma antigens. Through test, samples were reacted with conjugated colours (anti-Toxoplasma of monoclonal mouse microsphere (red antibodies)), the samples were dried before that, the combination then travelled to reach membranes via the act of capillaries. While samples move via the membranes tests, tinted particle were migrated. In positive results, certain antibodies that have existed on the membranes captured these particles which lead to appearance of red tinted line that can clearly observed while the other result appears in a green tinted line (the negative results that represent the control samples).

IgM Toxoplasma antibodies and IgG determination

To achieve the qualitative and quantitative investigation to IgG Toxoplasma gondii and IgM, two kinds of kits were used to detect the antibodies in infected women serums; these kits are (Toxoplasma IgM, EIA, enzyme immunoassay) and Toxoplasma IgG EIA enzyme immunoassay) (ACON Laboratories, Inc. San Diego, USA). Toxoplasma antibodies levels were assessed using ELISA technique. The level of IgM or IgG below 0.9 IU/ml was considered negative and from 0.9 to 0.99 IU/ml was equivocal limit and should be rechecked, while positive level was equal to 1.0 IU/ml or above.

Quantitative estimation of the complement:

Specimens for patients and healthy women for complement (C3,C4) were done by radioimmuno diffusion plate (LTA, Italy) according to method of Mancini, 1965 [22].

Estimation of the level of IL-6, IL-17 and Dopamine

The levels of IL-6, IL-17 and dopamine were estimated in 50 patients who were found anti-Toxoplasma antibodies positive according to manual procedure of Cusabio Biotech (China).

Statistical analyses

The results were analyzed using Statistical Package for the Social Sciences (SPSS) version-18 (T-test).

RESULTS

Diagnosis of *T. gondii*

The anti-Toxoplasma IgM present in 14 cases with a percent 10.37 %,also , the anti-Toxoplasma IgG present in 26 cases with a percent of 19.25 % while both IgM and IgG present in only 10 cases with a percent of 20% out of 135 cases in Immunochromatography method (Table-1).

Table-1: Distribution of anti-Toxoplasma gondii IgG and IgM antibodies using immunochromatography method in women with repeated abortion

Anti-Toxoplasma antibodies	Total	Positive		Negative	
		No.	%	No.	%
IgM	135	14	10.37	124	89.63
IgG	135	26	19.25	109	80.75
IgM +IgG	135	10	7.40	125	92.60

The level of anti-Toxoplasma IgM present in 50 cases was 1.57 in comparison with healthy control 0.27 also, the level of anti-Toxoplasma IgG was 142.73 in comparison with healthy control 14.66 (Table-2).

Table-2: The Level of anti-Toxoplasma gondii IgM antibodies using ELISA method in infected women

Treatment	Total	IgM IU/ml	
		Mean	SD
Infected women	50	1.57	1.64
Healthy Control	30	0.27	0.09

Table-3: The Level of anti-Toxoplasma gondii IgG antibodies using ELISA method in Infected women

Treatment	Total	IgG IU/ml	
		Mean	SD
Infected women	50	142.73	94.66
Healthy Control	30	14.66	5.92

The level of C3 and C4 increased significantly in patients groups in comparison with healthy control (Tables 4 and 5).

Table-4: The Level of C3 Complement Infected women and healthy control

Treatment	Total	C3 mg/dl	
		Mean	SD
Infected women	50	148.6	55.7
Healthy Control	30	129.7	13.6

Table-5: The Level of C4 Complement Infected women and healthy control

Treatment	Total	C4 mg/dl	
		Mean	SD
Infected women	50	26.7	13.3
Healthy Control	30	23.2	4.8

The level of IL-6 pg/ml increased significantly in patients groups in comparison with healthy control (Table-6).

Table-6: The Level of Interleukin-6 in Infected women and healthy control

Treatment	Total	IL-6 Pg/ml	
		Mean	SD
Infected women	50	3.22	1.61
Healthy Control	30	1.78	0.68

The level of IL-17 pg/ml increased significantly in patients groups in comparison with healthy control (Table-7).

Table-7: The Level of Interleukin-17 in Infected women and healthy control

Treatment	Total	IL-17 Pg/ml	
		Mean	SD
Infected women	50	3.26	1.69
Healthy Control	30	1.61	0.69

The level of Dopamine ng/ml increased significantly in patients groups in comparison with healthy control (Table-8).

Table-8: The Level of Dopamine in Infected women and healthy control.

Treatment	Total	Dopamine ng/ml	
		Mean	SD
Infected women	50	1.95	0.86
healthy Control	30	1.03	0.33

DISCUSSION

The majority of acquired infection in healthy individual are benign and either asymptomatic or with vague symptoms. About one-third of the world's population is estimated to carry Toxoplasma infection. There are large variations in prevalence and within different countries in humans [23]. The result indicated Anti –Toxoplasma IgG was 26 Cases while IgM was 14 cases in immunochromatography method (Table-1) while the level of anti-Toxoplasma IgG was (142.73) and IgM (1.57) by ELISA method (Table 2 & 3) in a general The prevalence of infection is related to several factors including nutritional habits, contact with soil, rural or urban settings and frequency of contact with domestic animals and climatic condition such as humidity and pregnancy [24]., The prevalence of anti- *T. gondii* antibody observed in the study was in agreement with seroprevalence data from previous studies conducted in our country 66 (29.2%) from a survey carried out in Salah– Adden government on 226 pregnant women had Toxoplasmosis. Anti-Toxoplasma IgG was 59 (26.1%) of cases while IgM antibody was 7 (3.1%) of cases [24]. The increasing in level of C3 and C4 (Table 4 & 5) may be contributed to the ability of the parasite to repair their membrane after the membrane attack complex of complement has been assembled on its surface and these factor may necessitate a high input of complement activity in order to cause cytolysis of the organism [25]. The increasing level of IL-6 (Table- 6) may be involved in early development of acute phase response, maintenance of hematopoi-esis [26] and immune barriers in ocular [27] as well as cerebral Toxoplasmosis [28]. It makes the NK cells to increment their cytotoxic activity and is also involved in the maturation of antibody secreting B lymphocytes and the differentiation of T lymphocytes [29]. Different immune cells such as MΦs, endothelial cells, monocytes, and fibroblasts are involved in the production of IL-6. This cytokine functions synergistically withIL-1β and TNF-α. Hence, it is regarded as a remarkable pyrogenic factor mediating dominantly the production of hepatocyte based acute in- flammatory proteins [30]. The significant increasing level of IL-17 (Table-7) are secreted from Th17 cells. But a variety of immune cells including CD8+ and NK cells [31, 32]. also secrete IL17. It is an inflammatory cytokine that provides innate immunity from the recruitment of neutrophils [33] which make the host resistant against *T. gondii* infection [34]. A subset of CD4+ T cells has been identified which produces IL-6 , IL-17A, IL-17F, and TNF in response to IL -23 [35, 36]. The significant increasing level of dopamine (Table-8) may be due to ability of *T. gondii* infection as a risk factor for the development of schizophrenia and depression in humans. As *T. gondii* forms cysts that are located in various anatomical sites including the brain during a chronic infection, it is well placed anatomically to mediate these effects directly. The *T. gondii* genome is known to contain 2 aromatic amino acid hydroxylases that potentially could directly affect dopamine and/or serotonin biosynthesis [37].

CONCLUSION

The result indicated presence of the anti-Toxoplasma IgG in 26 cases , Anti-Toxoplasma IgM in 14 cases and 10 in both out of 135 cases of aborted women with Toxoplasma in immunochromatography methods while the level of IgM 1.57 IU/ml and IgG 142.73. also, the result indicated significant increasing levels of IL-6 and IL-17 ,Complement and dopamine in patients sera in comparison with healthy control.

REFERENCES

1. Navia, B. A., Petito, C. K., Gold, J. W., Cho, E. S., Jordan, B. D., & Price, R. W. (1986). Cerebral toxoplasmosis complicating the acquired immune deficiency syndrome: clinical and neuropathological findings in 27 patients. *Annals of neurology*, 19(3), 224-238.
2. Pappas, G., Roussos, N., & Falagas, M. E. (2009). Toxoplasmosis snapshots: global status of Toxoplasma gondii seroprevalence and implications for pregnancy and congenital toxoplasmosis. *International journal for parasitology*, 39(12), 1385-1394.
3. Dunay, I. R., DaMatta, R. A., Fux, B., Presti, R., Greco, S., Colonna, M., & Sibley, L. D. (2008). Gr1+ inflammatory monocytes are required for mucosal resistance to the pathogen Toxoplasma gondii. *Immunity*, 29(2), 306-317.
4. Mordue, D. G., & Sibley, L. D. (2003). A novel population of Gr-1+-activated macrophages induced during acute toxoplasmosis. *Journal of leukocyte biology*, 74(6), 1015-1025.
5. Bliss, S. K., Butcher, B. A., & Denkers, E. Y. (2000). Rapid recruitment of neutrophils containing prestored IL-12 during microbial infection. *The Journal of Immunology*, 165(8), 4515-4521.
6. Tait, E. D., Jordan, K. A., Dupont, C. D., Harris, T. H., Gregg, B., Wilson, E. H., ... & Hunter, C. A. (2010). Virulence of Toxoplasma gondii is associated with distinct dendritic cell responses and reduced numbers of activated CD8+ T cells. *The Journal of Immunology*, 185(3), 1502-1512.

7. Dunay, I. R., Fuchs, A., & Sibley, L. D. (2010). Inflammatory monocytes but not neutrophils are necessary to control infection with *Toxoplasma gondii* in mice. *Infection and immunity*, 78(4), 1564-1570.
8. Del Rio, L., Bennouna, S., Salinas, J., & Denkers, E. Y. (2001). CXCR2 deficiency confers impaired neutrophil recruitment and increased susceptibility during *Toxoplasma gondii* infection. *The Journal of Immunology*, 167(11), 6503-6509.
9. Liu, C. H., Fan, Y. T., Dias, A., Esper, L., Corn, R. A., Bafica, A., ... & Aliberti, J. (2006). Cutting edge: dendritic cells are essential for in vivo IL-12 production and development of resistance against *Toxoplasma gondii* infection in mice. *The Journal of Immunology*, 177(1), 31-35.
10. Gazzinelli, R. T., Hieny, S., Wynn, T. A., Wolf, S., & Sher, A. (1993). Interleukin 12 is required for the T-lymphocyte-independent induction of interferon gamma by an intracellular parasite and induces resistance in T-cell-deficient hosts. *Proceedings of the National Academy of Sciences*, 90(13), 6115-6119.
11. Hunter, C. A., Subauste, C. S., Van Cleave, V. H., & Remington, J. S. (1994). Production of gamma interferon by natural killer cells from *Toxoplasma gondii*-infected SCID mice: regulation by interleukin-10, interleukin-12, and tumor necrosis factor alpha. *Infection and immunity*, 62(7), 2818-2824.
12. Gazzinelli, R. T., Wysocka, M., Hayashi, S., Denkers, E. Y., Hieny, S., Caspar, P., ... & Sher, A. (1994). Parasite-induced IL-12 stimulates early IFN-gamma synthesis and resistance during acute infection with *Toxoplasma gondii*. *The Journal of Immunology*, 153(6), 2533-2543.
13. Suzuki, Y., Orellana, M. A., Schreiber, R. D., & Remington, J. S. (1988). Interferon- γ : the major mediator of resistance against *Toxoplasma gondii*. *Science*, 240(4851), 516-518.
14. Bliss, S. K., Marshall, A. J., Zhang, Y., & Denkers, E. Y. (1999). Human polymorphonuclear leukocytes produce IL-12, TNF- α , and the chemokines macrophage-inflammatory protein-1 α and-1 β in response to *Toxoplasma gondii* antigens. *The Journal of Immunology*, 162(12), 7369-7375.
15. Bliss, S. K., Gavrilescu, L. C., Alcaraz, A., & Denkers, E. Y. (2001). Neutrophil depletion during *Toxoplasma gondii* infection leads to impaired immunity and lethal systemic pathology. *Infection and immunity*, 69(8), 4898-4905.
16. Johnson, L. L., VanderVegt, F. P., & Havell, E. A. (1993). Gamma interferon-dependent temporary resistance to acute *Toxoplasma gondii* infection independent of CD4+ or CD8+ lymphocytes. *Infection and immunity*, 61(12), 5174-5180.
17. Sher, A., Oswald, I. P., Hieny, S., & Gazzinelli, R. T. (1993). *Toxoplasma gondii* induces a T-independent IFN-gamma response in natural killer cells that requires both adherent accessory cells and tumor necrosis factor-alpha. *The Journal of Immunology*, 150(9), 3982-3989.
18. Denkers, E. Y., Gazzinelli, R. T., Martin, D., & Sher, A. (1993). Emergence of NK1.1+ cells as effectors of IFN-gamma dependent immunity to *Toxoplasma gondii* in MHC class I-deficient mice. *The Journal of experimental medicine*, 178(5), 1465-1472.
19. Hauser Jr, W. E., Sharma, S. D., & Remington, J. S. (1982). Natural killer cells induced by acute and chronic toxoplasma infection. *Cellular immunology*, 69(2), 330-346.
20. Kang, H., & Suzuki, Y. (2001). Requirement of non-T cells that produce gamma interferon for prevention of reactivation of *Toxoplasma gondii* infection in the brain. *Infection and immunity*, 69(5), 2920-2927.
21. Combe, C. L., Curiel, T. J., Moretto, M. M., & Khan, I. A. (2005). NK cells help to induce CD8+-T-cell immunity against *Toxoplasma gondii* in the absence of CD4+ T cells. *Infection and immunity*, 73(8), 4913-4921.
22. Mancini, H. S., Carbonara, A. O., & Heremans, J. F. (1956). *Immunochemistry*, 2, 235-21.
23. Torda, A. (2001). Toxoplasmosis: are cats really the source?. *Australian Family Physician*, 30(8), 743-747.
24. ADdory, A. Z. R. A. (2011). Seroepidemiological study of Toxoplasmosis among pregnant women in Salah-Adden government. *Tikrit Medical Journal*, 17(1), 64-73.
25. Shani, W. S., Bushra, H. S., & Nabel, E. W. (2012). Levels of Immunoglobulins and complements in sera of patients with toxoplasmosis. *Basrah Journal of Scienec (B) Vol*, 30(1), 72-77.
26. Akira, S., Taga, T., & Kishimoto, T. (1993). Interleukin-6 in biology and medicine. *Advances in immunology*, 54, 1-78.
27. Lyons, R. E., Anthony, J. P., Ferguson, D. J. P., Byrne, N., Alexander, J., Roberts, F., & Roberts, C. W. (2001). Immunological studies of chronic ocular toxoplasmosis: up-regulation of major histocompatibility complex class I and transforming growth factor β and a protective role for interleukin-6. *Infection and immunity*, 69(4), 2589-2595.
28. Suzuki, Y., Yang, Q., Conley, F. K., Abrams, J. S., & Remington, J. S. (1994). Antibody against interleukin-6 reduces inflammation and numbers of cysts in brains of mice with toxoplasmic encephalitis. *Infection and immunity*, 62(7), 2773-2778.
29. Filisetti, D., & Candolfi, E. (2004). Immune response to *Toxoplasma gondii*. *Ann Ist Super Sanita*, 40(1), 71-80.
30. Lockhart, E., Green, A. M., & Flynn, J. L. (2006). IL-17 production is dominated by $\gamma\delta$ T cells rather than CD4 T cells during *Mycobacterium tuberculosis* infection. *The Journal of Immunology*, 177(7), 4662-4669.
31. Michel, M. L., Keller, A. C., Paget, C., Fujio, M., Trottein, F., Savage, P. B., ... & Leite-de-Moraes, M. C. (2007). Identification of an IL-17-producing NK1.1neg iNKT cell population involved in airway neutrophilia. *The Journal of experimental medicine*, 204(5), 995-1001.

32. Rachitskaya, A. V., Hansen, A. M., & Horai, R. (2008). Cutting edge: NKT cells constitutively express IL-23 receptor and ROR γ and rapidly produce IL-17 upon receptor ligation in an IL-6-independent fashion. *J Immunol*, 180(8), 5167–5171.
33. Xue, M. L., Thakur, A., & Willcox, M. (2002). Macrophage inflammatory protein-2 and vascular endothelial growth factor regulate corneal neovascularization induced by infection with *Pseudomonas aeruginosa* in mice. *Immunology and cell biology*, 80(4), 323-327.
34. Kelly, M. N., Kolls, J. K., Happel, K., Schwartzman, J. D., Schwarzenberger, P., Combe, C., ... & Khan, I. A. (2005). Interleukin-17/interleukin-17 receptor-mediated signaling is important for generation of an optimal polymorphonuclear response against *Toxoplasma gondii* infection. *Infection and immunity*, 73(1), 617-621.
35. Aggarwal, S., Ghilardi, N., Xie, M. H., de Sauvage, F. J., & Gurney, A. L. (2003). Interleukin-23 promotes a distinct CD4 T cell activation state characterized by the production of interleukin-17. *Journal of Biological Chemistry*, 278(3), 1910-1914.
36. Langrish, C. L., Chen, Y., Blumenschein, W. M., Mattson, J., Basham, B., Sedgwick, J. D., ... & Cua, D. J. (2005). IL-23 drives a pathogenic T cell population that induces autoimmune inflammation. *The Journal of experimental medicine*, 201(2), 233-240.
37. Henriquez, S. A., Brett, R., Alexander, J., Pratt, J., & Roberts, C. W. (2009). Neuropsychiatric disease and *Toxoplasma gondii* infection. *Neuroimmunomodulation*, 16(2), 122-133.