SAR Journal of Pathology and Microbiology

Abbreviated Key Title: *SAR J Pathol Microbiol* Home page: <u>https://sarpublication.com/journal/sarjpm/home</u> DOI: https://doi.org/10.36346/sarjpm.2025.v06i04.003



Original Research Article

Anti-Microbial, Anti-Diabetic (α-glucosidase and α-amylase) Inhibitory Potential and Screening of Bioactive Chemical Compounds of Coriandrum Sativum Seeds Extract

Dr. Bashar Oda Jawad^{1*}, Dr. Abeer Fauzi Murad Al-Rubaye²

¹Hammurabi College of Medicine, University of Babylon, Iraq ²Department of Biology, College of Science for Women, University of Babylon, Iraq

*Corresponding Author: Dr. Bashar Oda Jawad

Hammurabi College of Medicine, University of Babylon, Iraq

Article History: | Received: 02.06.2025 | Accepted: 18.07.2025 | Published: 19.07.2025 |

Abstract: Background: Secondary metabolites, or SMs, are substances that are necessary for an organism's (cell's) survival yet have an impact on the organism's surroundings. These substances are important for protecting plants from biotic and abiotic stressors. Secondary metabolites are members of the other metabolite families and can be highly generated in response to stress. By participating in nutrition and reproduction, primary metabolites play a crucial role in providing metabolic activities. Accordingly, a small number of SMs are particularly chemical, such as medications, tastes, scents, insecticides, and dyes, and are consequently very cost-effective. Purposes: The current study's objectives are: Evaluation of the extract from Coriandrum sativum seeds' antimicrobial and anti-diabetic (a-glucosidase and aamylase) inhibitory properties as well as the identification of its bioactive chemical components. Materials and Methods: In the Hillah secondary city's local market, the coriander seeds were ground into a powder. It is ground to expand the surface area and speed up the extraction process. Glass beads and sample were combined in a 2:1 v/v ratio for the experiment. One hundred grammes of the sample was put into the glass-bead extraction jar, and it was extracted for around four hours, or until the extraction was finished. To make sure the extraction process was completed, the extract was harvested every 30 minutes, and the obtained oil was weighed right away. *Results:* Peak Wave number cm-¹, Intensity, Type of Intensity, Bond, Type of Vibration, and Functional group assignment were as follows: (667.37, 67.245, Strong, =C-H, Alkenes), (794.67, 77.258, Strong, =C-H, Alkenes), (1045.42, 65.642, Strong, C-F, alkyl halides), (1238.30, 81.459, Strong, C-F, alkyl halides), (1519.91, 75.457, Medium, C=C, Aromatic), (1531.48, 74.327, Medium, C=C, Aromatic), (1633.71, 68.420, Bending, N-H, Amide), (2854.65, 86.016, Strong, C-H, Alkane), (2924.09, 81.209, Strong, C-H, Alkane), (3163.26, 84.032, Strong, C-H, Alkane). Based on the extract type (Crude methanolic extract, ethanolic fraction, and standard acarbose), the corresponding inhibitory potencies against α -amylase were 89.73 ± 0.54 , 61.10 ± 0.27 , and 17.30 ± 0.09 , respectively. 63.25 ± 0.29 , 31.83 ± 0.15 , and 15.89 ± 0.06 were the measured inhibitory potencies against α -glucosidase activity, respectively. At a significantly (P < 0.05) lower concentration, crude methanolic extract demonstrated a better percentage inhibition against α -amylase than the antidiabetic medication acarbose (72.16%).

Keywords: Anti-Microbial, Anti-Diabetic, Inhibitory Potential, Coriandrum Sativum, FTIR, Secondary Metabolites. Copyright © 2025 The Author(s): This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International License (CC BY-NC 4.0) which permits unrestricted use, distribution, and reproduction in any medium for non-commercial use provided the original author and source are credited.

INTRODUCTION

These days, tens of thousands of diseases attack us every day, and in order to combat these illnesses, we have discovered that it is more important to find natural antibiotics that work well against the influx of illnesses that could endanger us due to infections with grampositive and gram-negative bacteria. Although there are currently synthetic antibiotics available in stores, each antibiotic has unique adverse effects. Scientists are therefore searching for natural antibiotics, particularly those derived from plants [1, 2]. The Umbelliferae family

Citation: Bashar Oda Jawad & Abeer Fauzi Murad Al-Rubaye (2025). Anti-Microbial, Anti-Diabetic (α -glucosidase and α -amylase) Inhibitory Potential and Screening of Bioactive Chemical Compounds of Coriandrum Sativum Seeds Extract. *SAR J Pathol Microbiol*, 6(4), 156-165.

includes the vertical annual herb Coriandrum sativum, which has a unique taproot with shooting up directions that vary from 20 to 70 cm. The Coriandrum sativum crops require 60 to 120 days to ripen and 45 to 60 days to reach the flower stage after being spread, depending on their variety [3]. At least three or ten umbels, each with ten or fifteen flowers, are produced at each Coriandrum branch. According to reports in [4, 5], the pollination and flowering biology of coriander differs from that of umbelliferous plants. A single umbel's flowering typically takes five to seven days to complete. However, the weather will determine how long it lasts. This delay is significantly increased in cold and rainy weather. Consequently, blooms that did not receive favourable climatic conditions would produce fewer fruits, or many fruits will only have one covering, such as a mericarp with a seed. Important cross-pollinating insects avoid visiting the flowers during cold or rainy weather [6, 7]. It follows that several insect species visit or pollinate the Coriandrum sativum umbels when the conditions are right. Insect species that are specific to the coriander's growing location pollinate it [8]. While none of the plants were incapable, it was discovered that 25% of the plants fertilised with other plants' pollen were capable of self-pollination [9]. This simple method, which uses the paternity plant's marker gene to provide information about the biology of plant pollination, is also interesting to consider in relation to plant breeding because it eliminates the laborious emasculation process and is applicable to crossings where caution is not needed. These authors also believed that even after three generations of self-pollinating, no signs of inbreeding depression were observed [10]. The plant known as Coriandrum sativum is a crossbred crop. Coriandrum Sativum, also referred to as Dhanya, is a highly valued medicinal ayurveda tree. Several parts of Coriandrum sativum have been shown to produce essential oil, fatty acids, flavonoids, and sterols. This plant's leaves, seeds, flowers, and fruit all have antifungal, antioxidant, antidiabetic, antihelmintic, anti-mutagenic soporifichypnotic, diuretic, anticonvulsant, cholesterol-lowering, anticancer, anxiolytic, hepatoprotective, anti-ulcer, antiprotozoal, defensive role against lead poisoning, heavy metal detoxification, and post-coital [11, 12], properties. Coriandrum sativum's dried fruits are useful for making most spices that retain their natural and beneficial effects in food, which benefits our digestive systems. Since ancient times, it has been essential for adding flavour to meat and baked goods. The study's goals are: Using Fourier Transform Infrared Spectroscopy (FTIR), screen the phytochemical and identify certain volatile components in the Coriandrum sativum ethanolic whole plant extract. Examine its antimicrobial properties. Alpha-amylase and alpha-glucosidase inhibitory action as an anti-diabetic.

MATERIALS AND METHODS

Preparing Samples

After being harvested and given time to dry, coriander seeds were pulverised in a blender. Before

being used, each sample powder was kept in a deep freezer in plastic bags. To extract the ethanolic fraction, 20 g of C. sativum was steeped in 100 ml of sterile distilled water in several sterile flasks. Over the course of two days, the flasks were periodically shaken. The contents of the flask were filtered.

Antimicrobial Properties

The first step was to dilute 6–7g of nutritional broth with 500 ml of water. After that, put it in the autoclave, pour this, and wait till the next day. Put a bacterial strain on it the following day; if it responds in any way, the result will be positive; if it does not, the result will be negative. To determine the antibacterial activity of the Coriandrum sativum seeds against the microorganisms, the existence of inhibitory zones, zone diameter, and MIC levels were employed. Apply a bacterial strain the following day if it produces any results; if not, the result will be negative; if it produces some results, the result will be positive.

Extract from Coriandrum Sativum Seeds Fourier Transform Infrared Spectroscopy (FTIR) Analysis

Determining how a given sample absorbs light at all wavelengths is the aim of any spectroscopy measurement, including FTIR and ultraviolet-visible (UV-Vis) spectroscopy. A technique used to obtain an absorption infrared spectrum of a solid, liquid, or gas is called Fourier transform infrared spectroscopy (FTIR). You may obtain excellent spectral resolution in an infinite spectrum domain using FTIR spectroscopy. Each of the GLVs' FTIR spectra were captured by running a variety of data that had previously been collected using an FTIR instrument and processed mostly in laboratories using computer-based software, nearly all of which are PC-based. A suitable and thin layer was created by physically compressing the combination under examination, and a small amount of laboratory-chopped leaves was subsequently converted to pellets using KBr in order to conduct the experiment before FTIR analysis. Information was actually gathered on the number of waves that made up 4000 cm 1 to 500 cm 1 in order to obtain accurate and thoroughly studied facts about the transfer of infrared light. All experimental samples underwent three different tests in this study, with KBr pellets that had not been treated serving as a control.

α-Amylase Inhibi

To demonstrate the extract's and the fractions' 0-amylase inactivating effect, a few minor modifications were made to the standard procedure. There were 20 millilitres of alpha amylase with two international units per millilitre, 200 millilitres of extract and fractions with varying concentrations of 0.5 milligrammes per millilitre, and 500 millilitres of 6.8 phosphate buffer with 100 millimolar phosphate. After 20 minutes, this was aliquoted in a 96-well plate and incubated at 37 C. At the incubation level, the preincubation temperature was 37 °C. After 30 minutes at 37 degrees Celsius, the mixture was once again placed in an incubator. As a substrate, 20 litres of 1% soluble starch in 100 mM phosphate buffer with a pH of 6.8 were added. After adding 100 litres of DNS colour reagent and stirring, the liquid was heated to a boil under constant pressure for ten minutes. To get the absorbance reading at 540 nanometres, a Multiplate Reader was then used. This was measured in order to calculate the final mixture's absorbance. Standard acarbose concentrations of 0.1 to 0.5 mg/ml were employed as control levels. One way to draw a comparison is to say that a material was synthesised at the same time that had been left to flower without any experimental manipulations (extracts and fractions). In particular, each experiment was replicated three times. The results were represented as a percentage of inhibition using the formula that was used. The IC50 value was determined by graphing the results of the enzyme activity inhibition by the various fraction values on graph paper.

The percentage of inhibition could be determined by applying the following formula:

% Inhibition = $(Abs_{control} - Abs_{extract}) / Abs_{control} \times 100$

Alpha Glucosidase Inhibitory Assay

Alpha glucosidase functions were analysed in order to determine the extract's and the fractions' inhibitory properties. In other instances, the analysis was conducted using the standard procedure with a few small modifications. In order to create serum samples, the sample was pre-cooled in a 96-well plate for 15 minutes at 37 degrees Celsius. The reaction mixture also contained 50 litres of the 6.8 phosphate buffer solution at a concentration of 100 mM, 10 litres of the purified alpha-glucosidase at one unit/mL, and 20 litres of 0.500 mg/mL of the nine distinct extracts and the first nine fractions. 37 degrees Celsius was used for the preincubation phase. Twenty litres of P-NPG with a five millimolar concentration were added as a substrate after the solution had been incubated for an additional twenty minutes at 37 degrees Celsius. This process was halted by adding 50 litres of 0.1 M sodium carbonate solution. With the aid of a multiplate reader, this experiment accurately estimated the absorbance of the recently released nitrophenol; the reading was computed at 405 nm. Acarbose was used as a control measurement in the interim and was detected in the sample at a concentration of 0.5 mg/mL. To compare the outcomes (readings), these three tests were conducted roughly three times. Additionally, a control experiment was conducted concurrently without the application of the study chemical. To ensure that the results of the studies were as precise as possible, they were carried out three times. To calculate the 100% inhibitory activity of 2-larabinose using 1/10 of the necessary quantity of 100% inhibitory activity, the following expression was used to calculate the percentage inhibition of 100%:

(Abscontrol - Absextract) = Inhibition Abscontrol changing to a percentage

In this case, the absorbance of the fractions and the control are denoted by the symbols a extract and A control, respectively. The fractional amounts required to limit enzyme activity to 50% were indicated by the IC50 values that were computed using the visual displays.

Analysis of Statistics

To ascertain whether parametric data was statistically significant, we used the Student t-test if the p-value was less than or equal to 0.05.

RESULTS AND DISCUSSION

Hundreds of thousands of low molecular weight organic syntheses are produced by the plant kingdom. The research community has divided these substances into three main groups based on their alleged actions: primary metabolites, which are directly necessary for plant growth; secondary (or specialized) metabolites, which participate in interactions between plants and their surroundings; and hormones, which regulate the body's functions and metabolism [13]. Over the years, plant biology theory and experimentation have been guided by this functional trichotomy of plant metabolism. However, there were never clear-cut biological boundaries between these several metabolite types. A recent wave of genetic and chemical research has further blurred the lines between these groups by demonstrating that secondary metabolites are also multifunctional, acting as strong regulators of plant growth and defence in addition to primary metabolites sensu lato (e.g., the monosaccharide ribose). Numerous adaptive circumstances, including effective signalling and inexpensive storage and recycling, could have supported this functional diversity of secondary metabolites. Given that adapted herbivores use plant secondary metabolites in a variety of ways that are similar to their functions in plants, the multifunctionality of secondary metabolites may provide new insights [14, 15], into the production of ontogenetic defences and enhance research on plantherbivore interactions. Bond = C--H, Type of Vibration Bending, Intensity 67.245, Type of Intensity 6 Strong, Peak Wave 667.37 cm-1', and Functional Group Assignment Alkenes, these values were equally distributed in the process of preparing the final FTIR analysis result: (794.67, 77.258, Strong, =C-H, Bending, Alkenes), (995.27, 67.253, Strong, =C-H, Bending, Alkenes), (1045.42, 65.642, Strong, C-F, Stretch, alkyl halides), (1238.30, 81.459, Strong, C-F, Stretch, alkyl halides), (1519.91, 75.457, Medium, C=C, Stretch, Aromatic), (1531.48, 74.327, Medium, C=C, Stretch,

Amide), (2854.65, 86.016, Strong, C-H, Stretch, Alkane), (2924.09, 81.209, Strong, C-H, Stretch, Alkane), (3163.26, 84.032, Strong, C-H, Stretch, Alkane). Secondary metabolites are components that are beneficial natural compounds that are produced by plants' secondary metabolism. The induction of morphological differentiation is linked to the synthesis of different secondary metabolites, and it is seen that as the plant grows, the induction of morphological differentiation and cell maturation occurs. When compared to non-differentiated or less-differentiated tissues, it is shown that the production of secondary metabolites of different tissues is noticeably higher in vitro. Such metabolites have many advantages, including rapid recovery of the products and easy selection of cell lines to increase the synthesis of secondary metabolites

in plants that are difficult to grow, costly, or resistant, such as plant cultures. Given the active growth of this line of inquiry, numerous further examples of plant metabolic engineering can be provided. According to this perspective, metabolic engineering is probably a significant step in the right direction, but tampering with genes is not the answer to all of the problems that have prevented the economic success of the secondary metabolite business in plants and New methods of cheaply and commercially growing even uncommon and exotic plants, their cells, and the compounds they produce could result from advancements in plant cell cultures [16-19]. In both plants and cultures, the biosynthesis pathways of desired molecules are frequently primitive, necessitating the creation of information at the cellular and molecular level.



Figure 1. Fourier-transform infrar	d spectroscopic profile solid analysis of	seed extract of Coriandrum sativum.

No.	Peak (Wave number cm- ¹)	Intensity	Corr. Intensity	Base (H)	Base (L)	Area	Corr. Area	Type of Intensity	Bond	Type of Vibration	Functional group assignment	Group frequency
2.	7 94.6 7	77.258	0.932	812.03	788.89	2.476	0.063	Strong	=C-H	Bending	Alkenes	650-1000
3.	995.27	67.253	3.724	1010.70	898.83	13.281	0.498	Strong	=C–H	Bending	Alkenes	650-1000
4.	1045.42	65.642	5.482	1132.21	1012.63	18.510	1.968	Strong	C-F	Stretch	alkyl halides	1000-1400
5.	1238.30	81.459	2.802	1265.30	1199.72	5.368	0.492	Strong	C-F	Stretch	alkyl halides	1000-1400
6.	1394.53	80.196	0.565	1398.39	1357.89	3.448	0.043	Strong	C-F	Stretch	alkyl halides	1000-1400
7.	1519.91	75.457	0.590	1521.84	1477.47	4.035	0.044	Medium	C=C	Stretch	Aromatic	1400-1600
8.	1531.48	74.327	0.914	1539.20	1521.84	2.192	0.054	Medium	C=C	Stretch	Aromatic	1400-1600
9.	1633.71	68.420	1.375	1637.56	1585.49	5.970	0.072	Bending	N-H	Stretch	Amide	1550-1640
10.	2854.65	86.016	3.390	2870.08	2812.21	2.751	0.223	Strong	С-Н	Stretch	Alkane	2850-3000
11.	2924.09	81.209	6.084	2951.09	2881.65	4.860	0.846	Strong	С-Н	Stretch	Alkane	2850-3000
12.	3163.26	84.032	0.163	3165.19	3134.33	2.174	0.010	Bending	N-H	Stretch	Amide	3100-3500













Crude methanolic extract and ethanolic fraction of coriandrum sativum seed extracts demonstrated 23.15±0.23 and 24.96±0.25 antibacterial activity against Enterobacter aerogenes, 17.05+0.16 and 21.94±0.20 against Staphylococcus saprophyticus, and 22.25±0.20 and 18.32 against Streptococcus pyogenes. The Coriandrum sativum seed extract fraction demonstrated the inhibition of 2-amylase and 2-glucosidase activities: Coriandrum sativum exhibited strong anti-2-amylase and anti-2-glucosidase properties, as seen in figures 6 and 7. According to the findings of the enzymatic inhibitor assay of the Coriandrum sativum fractions, the inhibitory activity was shown to be both phase and dose dependent. The study's highest dose exhibited the fastest inhibition. while its lowest concentration demonstrated the least amount of inhibition. Figure 6 shows the respective inhibitory potentials of crude methanolic extract, ethanolic fraction, and acarbose (standard) against aamylase, which were 89.73±0.54, 61.10±0.27, and 17.30±0.09. Following recording, inhibitory potency 0-glucosidase activity against was measured (63.25±0.29, 31.83±0.15, and 15.89±0.06), respectively, as shown in Figure 7. At a substantially lower concentration, the methanol and water fraction demonstrated superior percentage inhibition of 94.32 and 94.3 percent against alpha-amylase compared to the antidiabetic medication acarbose (72.16 percent) (P < 0.05). Mostly utilised as a seed crop, coriander is extensively distributed. The seed of the corriandrum sativum plant is primarily responsible for its therapeutic use. It is used as a suppository to treat rheumatism, worms, indigestion, and joint discomfort. The presence of hypoglycemic actions and the metabolism of carbohydrates have also been validated by the current investigation. It has been discovered that vital oil seeds and leaves contain explosive components that prevent the growth of microorganisms in a wide range of lipid peroxidation stores. Essential oil extracted from Coriandrum sativum leaves shown antibacterial action against both Gram-positive and Gram-negative bacteria. Given that it has been used for decades without showing any symptoms of toxicity, the plant in question has been shown to be safe [20-23]. Coriandrum sativum leaf essential oil has been shown to have antimicrobial properties. Furthermore, this toxicity was examined in order to ascertain whether C. sativum essential oil might be used individually as a phytotherapeutic product in relation to the Artemia salina mortality test [24, 25]. Two isolates of Candida albicans and 186 isolates of various Gram-positive bacterial populations obtained by isolating urine specimens were subjected to Coriander sativum fermentations and drug decoctions. The study looked at essential oil that was obtained from Coriandrum sativum L. (Apiaceae) leaves by hydrodistillation. The primary constituents were n-decanol (13.6 percent), 2E-decenal (15.9 percent), 2E-decen-1-ol (14.2 percent), and decanalling (14.3 percent). Dodecanal (4.36%), undecanol (3.37%), undecanal (3.23%), 2E-tridecen-1-al (6.75%), and 2Edodecenal (6.23%) are other components present in similarly

acceptable proportions. Compared to seeds, Coriandrum sativum leaves exhibited a greater antioxidant capacity, and the highest activity was obtained from methylated extract at equimolecular concentrations. Additionally, coriander had strong inhibitory effects against E. coli as well as other bacteria and fungi. Vegetable secondary metabolites called polyphenols or phenolic compounds have produced a wide range of complex phytochemicals with antioxidant activity and positive physiological effects. Numerous sources, including sage, rosemary, thyme, pepper, and oregano, have been shown to contain phenolic materials, which, in addition to flavonoids, also contain phenolic acids and have been shown to have significant antioxidant properties. These compounds can be found all over the plant kingdom at any given time. Plants that grow coriander are well suited to tropical and subtropical climates. Fertile soil with tolerable organic matter, sunlight, and unrestricted water consumption is ideal for its growth. In general, it has been characterised cholesterol-lowering, anti-inflammatory, as and antidiabetic. In addition, it is used as a stomachic, aphrodisiac, anthelmintic, stimulant, analgesic, diuretic, painkiller, and hypoglycemic medication. Along with caffeic and chlorogenic acids, Coriandrum sativum also includes active phenolic acid complexes [26-30]. Additionally, the study reveals that the opposing oils found in C. sativum plant leaves may have antibacterial qualities against food-tolerated infections such as Salmonella species. High levels of antibacterial complexes with frequently advantageous properties are found in plant sources. Which not only effectively treat a variety of infectious disorders but also lessen the plethora of adverse effects typically linked to antimicrobial compounds. Applications unquestionably demonstrated the effectiveness of these natural pesticides. The fact that SM decomposes readily in plants and even in soil is a benefit, but it is also a drawback because synthetic pesticides are more lasting. Furthermore, compared to biopesticides, conventional pesticides are typically more effective. Conversely, plants are relatively simple to grow, and farmers without access to Western synthetic pesticides may be able to obtain sustainable plant yields through the use of biopesticides [31-33]. There will be several obstacles in the way of the creation of biorational pesticides because criminal laws do not favour different combinations of substances to be employed as pesticides. Natural substances are an undeveloped substitute, though. Because of these many applications, there is currently a global market for plant extracts and isolated SM that is valued at over \$10 billion USD annually. Therefore, it is a complicated problem that the biotechnologist must solve in order to come up with ways to produce the molecules in sufficient quantities and quality [34, 35]. This is the most popular and traditional approach, which entails growing the corresponding plants in the field or in greenhouses and then extracting their contents. Certain species produce higher quality and yields when new kinds are chosen. Cell and organ culture is an important in vitro propagation technique at that point. Direct impact in certain instances, genetic engineering of secondary metabolism has already been applied directly. For instance, in the case of Atropa belladonna, new plants with scopolamine as the primary product were produced after the plants were modified with the gene encoding the enzymes that convert L-hyoscyamine to Lscopolamine. Otherwise, genetics has frequently altered the victim's metabolism, resulting in plants with different ellesdesqueles and illettrefille colours [36-39]. The genes involved in the biosynthetic pathways must be identified and expressed, either in transgenic microorganisms or plants, for further study. If the endeavour is successful, recombinant bacteria or yeasts may be cultivated in the future, which would result in the production of useful plant SM. Combinatorial biosynthesis might then be an uncharted territory. Such a strategy has already proven successful and is based on genes that code for the enzymes that create antibiotics. Additionally, it has sparked a renewed interest in the regulation of SM synthesis as well as the location and mechanism of these compounds' sequestration inside the plant [40-43]. Alkaloid biosynthesis genes have been attempted to be inserted into microorganisms in recent years. Finally, using recombinant bacteria or yeast to produce useful alkaloids is a possibility. Genetic transfection of susceptible crop organisms may be another abundant source of exploitation in cases where the corresponding SM (whether of plant or microbial origin) is resistant to insects or diseases. Researchers from all over the world who work with cell or organ culture tried to find useful SM more than 20 years ago. Differentiated organ cultures, such as altered root cultures, are frequently almost as active as the whole plant, although undifferentiated cell cultures have frequently failed to produce such a chemical in reasonable yields [44]. Gene expression that is specific to cells and tissues appears to control these processes.

CONCLUSION

Not to mention the numerous additional compounds found in the plant kingdom and their wildly diverse distribution, we have only covered a small number of the secondary metabolites of plants in this study, along with their potential use in defensive mechanisms and ecological adaptability. Although they are not essential to the plant's life, the byproducts of secondary metabolism aid in the growth and development of the plant. Because secondary metabolites are essential to plants' ecological defences against herbivores and microbiological diseases, plants are also secondary metabolites. They also serve as pollinators and animal attractants (taste, colour, and odour) that spread seeds. They function as moderators of plant-microbe symbionts and plant-plant antagonistic interactions. Thus, the competition and survival of the plants are significantly influenced by the ecological roles of the secondary metabolites of the plants. The process engineering is another of genetic wav that biotechnological techniques are used to produce secondary metabolites. The same may also be

significantly influenced by plant tissue culture. The development of germs that are resistant to antibiotics has put natural goods in a precarious position to stop the decline in human health. The Coriandrum sativum plant is used to treat certain medications, and it can offer some important platforms for the development of safe, effective, and affordable medications. Bioactive phytochemical substances were found in the ethanolbased extract of Coriandrum sativum. The investigation of Coriandrum sativum using GC-MS revealed over twenty bioactive natural chemicals. Peak area, retention duration, molecular weight, and molecular formula are used to identify bioactive chemical substances. Thirteen functional groups—16 Peak (Wave number cm-1), Type of Intensity, Bond, and Functional group assignmentwere used in the FTIR analysis preparation. Bioactive secondary metabolites of Coriandrum sativum were found to have remarkable anti-Staphylococcal action.

REFERENCES

- 1. Matasyoh. Chemical composition and antimicrobial activity of the essential oil of Coriandrum sativum. Food Chemistry. 2009;113(2):526-529.
- Sourmaghi. Comparison of essential oil composition and antimicrobial activity of Coriandrum sativum L. extracted by hydrodistillation and microwaveassisted hydrodistillation. Journal of Food Science and Technology. 2015;52(4):2452-2457.
- Lo Cantore. Antibacterial activity of Coriandrum sativum L. and Foeniculum vulgare Miller var. vulgare (Miller) essential oils. Journal of Agricultural and Food Chemistry. 2004;52(26)7862-7866.
- 4. Begnami. Antimicrobial potential of Coriandrum sativum L. against different Candida species in vitro. Food chemistry. 2010;118(1):74-77.
- Methawiriyasilp. Preliminary study of antimicrobial activities on medicinal herbs of thai food ingredients. In III WOCMAP Congress on Medicinal and Aromatic Plants-Volume 1: Bioprospecting and Ethnopharmacology. 2003;675:111-114.
- Aftab A, Haider M, Ali Q, Malik A. Genetic evaluation for morphological traits of Coriandrum sativum grown under salt stress. Biological and Clinical Sciences Research Journal. 2021;2021(1):e006.
- Zare-Shehneh. Biological activities of a new antimicrobial peptide from Coriandrum sativum. Int J Biosci. 2014;4(6):89-99.
- Hassanen. Antioxidant and antimicrobial activity of celery (Apium graveolens) and coriander (Coriandrum sativum) herb and seed essential oils. Int. J. Curr. Microbiol. App. Sci. 2015;4(3):284-296.
- 9. Zardini. Analysis of antibacterial and antifungal activity of crude extracts from seeds of Coriandrum sativum. Gomal Journal of Medical Sciences. 2012;10(2).
- 10. Bilal, M, Nasir I, Tabassum B, Akrem A, Ahmad A, Ali, Q. Cytotoxicity and in-vitro antiviral activity of

lectin from Crocus vernus l. against potato virus Y. Applied Ecology and Environmental Research 2020;18:1301-1315.

- 11. Senthilkumar. Coriandrum sativum mediated synthesis of silver nanoparticles and evaluation of their biological characteristics. Materials Research Express. 2018;5(5):055032.
- Duman. Evaluation of bioactivity of linaloolrich essential oils from Ocimum basilucum and Coriandrum sativum varieties. Natural Product Communications. 2010;5(6): 1934578X1000500634.
- Malik A, Hafeez K, Nazar W, Naeem, M., Ali I, Ali Q, Ahmed Mujtaba Z, Rana M Hafeez M. Assessment of controversial risk factors in development of breast cancer: a study from local population. Biological and Clinical Sciences Research Journal. 2021;2021(1), e003.
- Burdock. Safety assessment of coriander (Coriandrum sativum L.) essential oil as a food ingredient. Food and Chemical Toxicology. 2009;47(1):22-34.
- Ahmad M, Ali Q, Hafeez M, Malik A. Improvement for biotic and abiotic stress tolerance in crop plants. Biological and Clinical Sciences Research Journal. 2021; 2021(1):e004.
- Nair. Comparative evaluation of Coriandrum sativum Linn. and Apium graveolens for antimicrobial activity. Research Journal of Pharmacy and Technology. 2017;10(2):541-544.
- 17. Ashebir. Evaluation of the antibacterial activity of crude preparations of Zingiber officinale (zingibl), Echinops spp. (kebericho), Coriandrum sativum (dimbilal) and Cymbopogan citratus (tej sar) on some food-borne pathogens. Ethiopian Journal of Health Sciences. 1999;9(1).
- Kačániová. Antioxidant, antimicrobial and antibiofilm activity of coriander (Coriandrum sativum L.) essential oil for its application in foods. Foods. 2020;9(3):282.
- Hameed B, Ali Q, Hafeez M, A M. Antibacterial and antifungal activity of fruit, seed and root extracts of Citrullus colocynthis plant. Biol Clin Sci Res J. 2020;2020(e033).
- 20. Mahendra. Coriandrum sativum: A daily use spice with great medicinal effect. Pharmacognosy Journal. 2011;3(21):84-88.
- 21. Halkier, B.A. and Gershenzon, J. (2006). Biology and biochemistry of glucosinolates. Annual Rev Plant Biol. 57: 303-333.
- 22. Kang, S.Y. and Kim, Y.C. (2007). Decursinol and decursin protect primary cultured rat cortical cells from glutamate-induced neurotoxicity. J Pharmacy Pharmacol. 59(6):863-870.
- Leustek, T. (2002). Sulfate metabolism. Somerville CR, Meyerowitz EM, eds, The Arabidopsis Book. American Society of Plant Biologists, Rockville, MD, doi/10.1199/ tab.0009.
- 24. Lipka, U., Fuchs, R., Kuhns, C., Petutschnig, E. and Lipka, V. (2010). Live and let die-Arabidopsis non-

Lipka, V. (2010). Live and let die-Atabidopsis in

host resistance to powdery mildews. Eur J Cell Biol. 89(2):194-199.

- 25. Mansfield, J.W. (2000). Antimicrobial compounds and resistances. The role of phytoalexins and phytoanticipins. In: slusarenko A. J., fraser R.S.S., vanloon L.C. and fraser R.S. (eds). Mechanism of resisyance to plant diseases. Springerverlag New York., pp325-363.
- Raskin, I., Ribnicky, D.M., Komarnytsky, S., Ilic, N., Poulev, A., Borisjuk, N., Brinker, A., Moreno, D.A. and Yakoby, R..N. (2002). Plant and human health in the twenty-first century. Trends Biotechnol. 20:522-531.
- Watson, A.A., Fleet, G.W.J., Asano, N., Molyneux, R.J. and Nash, R.J. (2001). Polyhydroxy latedalkaloid –natural occurrence and therapeutic applications. Phytochemistry. 56:265-295.
- Reddy, L., Odhav, B. and Bhoola, K.D. (2003). Natural product for cancer prevention: global perspective. Pharmacol Theraput. 99:1-13.
- Ncube, N.S., Afolayan, A.J. and Okoh, A.I. (2008). Assessment techniques of antimicrobial properties of natural compounds of plant origin: current methods and future trends. African J Biotechnol. 7 (12):1797-1806
- Tholl, D. (2006). Terpene Synthases and the regulation, diversity and biological roles of terpene metabolism. Curr Opinion Plant Biol. 9:297-304
- Shalaka, D.K. and Sandhya, P. (2009). Micropropagation and organogenesis in Adhatoda vasica for the estimation of vasine. Pharmacog Magaz. 5:539-363
- Lee, S.Y., Xu, H., Kim, Y.K. and Park, S.U. (2007). Rosmarinic acid production in hairy root cultures of Agastache rugosa Kuntze. World J Microbiol Biotechnol. 20:969-972
- 33. Krolicka, A., Kartanowicz, R., Wosinskia, S., Zpitter, A., Kaminski, M. and Lojkowska, E. (2006). Induction of secondary metabolite production in transformed callus of Ammi majus L. grown after electromagnetic treatment of the culture medium. Enz Microb Technol. 39: 1386 -1389.
- 34. Olivira, A.J.B., Koike, L., Reis, F.A.M. and Shepherd, S.L.K. (2001). Callus culture of Aspidosperma ramiflorum Muell.-Arg.: growth and alkaloid production. Acta Scientia. 23:609-612
- Sujanya, S., Poornasri, D.B. and Sai, I. (2008). In vitro suspension cultures of Azadirachta indica. J Biosci. 33:113-120
- 36. Wagiah, M.E., Alam, G., Wiryowidagdo, S. and Attia, K. (2008). Imporved production of the indole alkaloid cathin-6-one from cell suspension cultures of Brucea javanica (L.) Merr. Ind J Sci Technol. 1:1-6
- Kusakari, K., Yokoyama, M. and Inomata, S. (2000) Enhanced production of saikosaponins by root culture of Bupleurum falcatum L. using two step control of sugar concentration. Plant Cell Rep. 19:1115-1120

- Nikolaeva, T.N., Zagoskina, N.V. and Zaprometov, M.N. (2009). Production of phenolic compounds in callus cultures of tea plant under the effect of 2,4-D and NAA. Russ J Pl Physiol. 56:45-49
- Umamaheswai, A. and Lalitha, V. (2007). In vitro effect of various growth hormones in Capsicum annum L. on the callus induction and production of Capsiacin. J Plant Sci. 2:545-551
- 40. Nazif, N.M., Rady, M.R. and Seif, M.M. (2000). Stimulation of anthraquinone production in suspension cultures of Cassia acutifolia by salt stress. Fitoterapia. 71:34-40
- 41. Shrivastava, N., Patel, T. and Srivastava, A. (2006). Biosynthetic potential of invitro grown callus cells

of Cassia senna L. var. senna. Curr Sci. 90:1472-1473.

- 42. Zhao, J., Zhu, W. and Hu, Q. (2001). Enhanced catharanthine production in Catharanthus roseus cell cultures by combined elicitor treatment in shake flasks and bioreactors. Enz Microb Technol. 28:673-681.
- Lee-Parsons, C.W.T. and Rogce, A.J. (2006). Precursor limitations in methyl jasmonate-induced Catharanthus roseus cell cultures. Plant Cell Rep. 25:607-612.
- 44. Ramani, S. and Jayabaskaran, C. (2008). Enhanced catharathine and vindoline production in suspension cultures of Catharanthus roseus by ultraviolet-B light. J Mol Signal. 3:9-14.