

## The Role of Epigenetic Drugs in Cancer Therapy

Dr. Hiba Sabah Jasim<sup>1\*</sup>

<sup>1</sup>Department of Microbiology, College of Medicine, University of Baghdad, Iraq

\*Corresponding Author: Dr. Hiba Sabah Jasim

Department of Microbiology, College of Medicine, University of Baghdad, Iraq

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**Abstract:** Epigenetics refers to heritable and dynamic alterations in the whole genes which present in the sequence of nucleic acids. It consider as concurrent reaction with enzymes and several molecular ingredients. Epigenetic changes can cause the incorrect start of coding genes, allowing tumor development. Epigenetic modifiers are becoming potential targets in numerous malignant tumor therapies since they are sensitive to foreign drugs. Different epigenetic medicines that were lately refined and implicated in clinical experiences using of epigenetic medicines solitary or together with immunotherapy and chemotherapy has yielded promising outcomes, containing advanced anti-cancer impact, overcoming therapy resistant, and stimulation of the immune system defense.

**Keywords:** Epigenetic alteration, malignant tumor, Epigenetic drugs.

## INTRODUCTION

Waddington defined epigenetics in as "the department of biology that investigates the causative agents that act with the genome, that result as a phenotype. After that, epigenetic has been used to a broad domain of biological methods as further guide emerged the heritable modifications to the genes occur separately of somatic cell changes, neglected of their discriminating state [1, 2].

Finetuned epigenetic alterations, which commonly include DNA methylation, histone, or chromatin posttranslational alterations, the same as no signaling RNAs organizations are required to maintain heritable changes that occur are maintained during multiple cellular biological methods with the same genetic information.

Failure of aepigenetic markers to be heritable can result in incorrect gene code initiation or inhibition, which can lead to pathological alterations, including malignant tumors.

Epigenetic alterations, include several mechanisms such as methylation of DNA, chromatin alteration, signaling ofRNAs organizations, are required to maintain heritable changes. Failure of epigenetic markers to be heritable can result in incorrect gene code initiation or inhibition, which can lead to pathological alterations, including malignant tumors [3].

A malignant tumor develops as a result of accumulated genetic change, epigenetic modifications, and environmental factors. Many researches have focused on describing the genetic landscape of malignant tumors, ranging from oncogene signaling ways to the transformation series in various malignant tumor subtypes. Epigenetic impacts, unlike genetic transformation, relate to alterations in gene coding that do not result in permanent changes ain the genomic sequence. They are used more frequently in malignant tumor cells because epigenetic changes are reversible and more easily controlled than genomic evolution.

Aside from the fundamental changes that occur in somatic cells, a malignant tumors landscape is created by a variety of causes, adding to the dimensional complexity. The tumor microenvironment is made up of cells and textures that form a niche for stromal chemicals to feed tumor cells. Recent epigenetic alterations are focusing on tumor cells-

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TME connections as well as malignant tumor cell proliferation. Epigenetic therapy is becoming a common malignant tumor treatment approach due to the importance of epigenetic organization in malignant tumors [4].

In this review, there is an envision more advanced sequencing technologies that would allow for epigenome mapping and precise epigenetic alterations in malignant tumor therapy as a monotherapy or in combination with other new therapies, epigenetic therapy may benefit malignant tumor patients, epigenetic alter in tumor progression, and the envisage a more advanced sequencing technologies that would allow for epigenome mapping and precise epigenetic alterations in malignant tumor therapy. The disadvantages and potential hazards of recent epigenetic treatments are also demonstrated. The function of epigenetics in the genesis and therapy of malignant tumors may be illuminated by this review. DNA and RNA methylation are epigenetic alterations processes. The three forms of epigenetic alterations that are regarded to be the fundamental mechanisms of organization during carcinogenesis/malignant tumor growth are histone modifications, non-signaling RNAs, and histone modifications. The most well-studied epigenetic process is DNA and RNA a methylation and demethylation of DNA methylation, which occurs predominantly in CpG islands, which are located in the a5' allow region of more than half of all human genes [5].It performs a variety of tasks, such as X inactivation of chromosome, embryonic development, genomic imprinting, epigenetic reprogramming, cell identity establishment, and lineage specification [6]. To silence genes, methyl groups from S-adenosyl methionine are covalently attached to the 5 position of the cytosine pyrimidine ring. The a5-methylcytosine structure can either prevent transcriptional agents from attaching to DNA binding sites, or it can recruit methyl-binding domain proteins to help remodel chromatin. With histone modifications, gene coding becomes more constrained. The three DNA methyl transferases that catalyze DNA methylation are dmtase1, dmtase3a, and dmtase3b. Dmtase1, the maintenance DNA methyl transferase, has a higher catalytic activity and is crucial for maintaining DNA methylation status during replication by preferentially methylating hemi methylated DNA [7]. Apart from replication, "de novo" methyl transferases, dmtase3a and dmtase3b, generate and maintain the precise DNA methylation status in the genome.

On the other side, DNA demethylation is a reversible process that restores silenced genes that have been damaged by Dmtase. TET type one, two and three are ten-eleven translocation methyl cytosine dioxygenases that may convert a5mC ato a5-ahydroxymethylcytosine and then further oxidize 5-hmC to 5-formylcytosine and 5-carboxylcytosine [9]. Homeostasis between demethylation and methylation of the genome occurs as a dynamic mechanism of gene coding in a variety of cells. N6-methyl adenosine RNA methylation is becoming a clinical problem in epigenetic mechanisms and malignant tumor biology. M6 A is more abundant around the stop codon, the 3'UTR, and lengthy exons within internal areas [10]. It has an effect on transcription, degradation, splicing, and translation, among other aspects of RNA processing [11]. Changes in m6 A are both reversible and dynamic, according to new research. METTL type three, fourteen, and sixteen with wilms tumor associated protein type one, RNA binding motifaprotein type fifteen, zinc finger CCCH-type containing 13, and aKIAA1429 are among the methyl transferase components known as "writers" required for the production of RNA m6 A [12]. While interactions between components of erasers and readers such as eukaryotic initiation factor type three, heterogeneous nuclear ribonucleoprotein protein family, and insulin-like growth factor two mRNA binding proteins family can be used to designate m6 A methylation [13]. DNA is bundled into a very compact structure coated with histoneoctamers in chromatin, resulting in nucleosomes and the "beads on a string" form, which helps limit DNAasequence accessibility.aEach histone octamer is made up of a tetramer with two copies of histone 2A and two copies of histone 2B, separated by dimers of histone type three and four. These histone proteins have a lengthy N-terminal tail that can be methylated, acetylated, ubiquitylated, phosphorylated, ADP, and biotinylated at specific amino acid residues. These PTMs have been used extensively to investigate acetylation and methylation of lysine residues on H3 and H4. Histone acetylation is based on the "charge neutralization model," which argues that the positive charge of lysine residues on H3/H4 facilitates tight packing of negatively charged DNA with histones. The insertion of an acetyl group can loosen chromatin's tight structure, allowing transcriptional agents' access for transcription [14].

Histone acetyl transferases and deacetylases are enzymes that catalyze the addition and removal of acetyl sets. Unlike histone acetylation, histone methylation serves a more complex purpose that is dependent on the targeted residues. H3K4/36/79 methylation, for example, is assumed to contribute to active transcriptional status, whereas methylation at aH3K9/27 and H4K20 is thought to represent repressive epigenetic marks [15]. They are catalyzed by several histone methyl transferases, the bulk of which feature a SET domain. For example, enhancer of zeste type two which is specific for H3K27 trimethylation, which suppresses transcription [16].aHistone demethylase modulates transcriptional activity by removing methyl groups from such markers. RNAs that don't make a signal Non-signaling RNAs influence gene expression and make up more thana70% of the human genome [17]. The two main varieties are small ncRNA (non-coding RNA with less than a200 nucleotides) and long ncRNA (non-coding RNA with more than 200 nucleotides). The well-studied small non-coding RNA is MiRNA, a highly conserved ssRNA with around 20 nucleotides. Previously considered "junk RNAs," they are crucial mediators in biological resilience, buffering mild perturbations and maintaining organism homeostasis. Over a60% of protein-signaling genes in humans are influenced by miRNA organization [18]. A growing number of studies have discovered miRNA mechanisms in virtually all forms of malignant

cancers. Lengthy noncoding RNAs are a diverse set of long transcripts originating from a variety of chromosomal locations. They can influence target areas in the nucleus or cytoplasm by acting as chromatin regulators, enhancers, ncRNA sponges, molecular scaffolds, and other properties. In terms of splicing, 5'-cap, and polyadenylation, ncRNAs are comparable to mRNAs, with the exception of newly discovered circular RNAs, which lack a 5' cap and poly-A tail [19].

The ability of ncRNAs and circRNAs to encode functional peptides with short open reading frames has recently been identified, complicating their operations [20]. Abnormal epigenetic modifications occur with the formation of malignant tumors. Within cancers, epigenetic alterations occur. Tumor progression is triggered by the accumulation of changes in the genome and epigenome, which results in tumor heterogeneity and adaptability. Two models have been developed to explain how cancers progress: The malignant tumor stem cell paradigm, also known as the hierarchical model, considers malignant tumor stem cells to be the source of oncogenic transformation, the stochastic or clonal evolution model proposes that non-CSCs eventually acquire the original oncogenic alteration [21]. The heavily methylated scenario occurred on the allows of glutathione S-transferase pi type one and other genes involved in cell cycle, cell metastasis, and apoptosis, such as aCDKN2A, TIMPS, and DAPK, according to studies on prostate cancer [22]. On the other hand, hypomethylation of oncogenes has been discovered in a variety of cancers, including aLY6K in glioblastoma [23], aSLC34A2 in papillary thyroid carcinoma [24], aRBBP6 in colorectal cancer [25], and so on. DNA methylation variation is also likely to act as a genome-wide regulatory mechanism, rather than targeting specific genes. According to a recent investigation of the genomic and epigenomic landscape, hyper- and hypo methylation occurred in the early preneoplastic phases of HCC, which was significantly linked to deorganization of malignant tumor-related genes. Apart from abnormal DNA methylation, malignant tumors that follow the CSC model, in which bidirectional inter conversions are required, do not exhibit unbalanced histone change. The activating aH3K4me3 and repressive aH3K27me3 bivalent histone marks were first used to distinguish embryonic stem cells [27]. On the other hand, some malignant malignancies partially recreate this bivalency by down regulating onco fetal genes in malignant tumor cells [28]. They are dimensionally complicated because abnormal epigenetic changes on chromosomes are expected to interact during tumor formation. According to this research, histone modifications can be altered in conjunction with abnormal DNA methylation or non-signaling RNAs. Minor histone changes could have an impact on other histone marks. H3K36me2 expansion was blocked by NSD2 overcode in multiple myeloma, for example, favoring aH3K27ac enrichment and interactions with other regulatory elements, initiating carcinogenic pathways [29].

Malignant tumor cells exhibit a wide range of ncRNA and RNA changes in addition to chromosome abnormalities. Hundreds of aberrant miRNAs have been discovered in nearly every type of malignant tumor thus far. Putative onco-miRNAs like micro RNA type 21, 155, 210, and 122 are commonly upregulated in malignant tumors and target TSGs to provide malignant tumor cells a competitive edge. Tumor suppressor miRNAs (such as micro RNA type thirty-four and type two hundred) have the opposite effect on malignant tumors. It's worth noting that several miRNAs have several functions, especially in malignant tumors. The miR-181 family, for example, has four members, amiR-181a through miR-181d, all of which have inconsistent coding in various solid malignant tumors, implying that they are all related.

It's worth mentioning that members of the amiR181 family can be found in several chromosome clusters and are sensitive to a variety of epigenetic modifications, suggesting that other epigenetic changes can influence miRNA synthesis. Interactions between distinct epigenetic mechanisms could also have a synergistic or antagonistic influence on genetic code. Another example is that the amiR-200 family can reduce EMT-related gene transcription by targeting aZEB1 and aZEB2 [31].

Non coding RNAs and circRNAs, as well as epigenetic influences on miRNA synthesis, can cause mature miRNAs to lose their functionality. It has long been known that ncRNAs and circRNAs act as miRNA sponges, attaching directly to miRNAs and thereby partially negating their effects [32]. According to a recent study in triple-negative breast malignant tumors, the ncRNA FAM83H-aAS1 sequestered miR-136-a5p, which is thought to decrease MTDH-induced proliferation, migration, and invasion of TNBC cells. The circRNA circFUT8 was down regulated and shown to be a tumor suppressor in bladder malignant tumors via the miR-570-a3p/KLF10 axis [34].

Finally, deorganization of m6 A alterations in both pri-miRNA and mRNA methoding have been discovered in several malignant tumors. The "writers" METTL3 and METTL14 have been found in abundance in a variety of malignant tumors, and they perform an oncogenic role by boosting mRNA translation via m6 A alteration [35]. They can also alter the methoding of pri-miRNA in HCC by recruiting DGCR8 in a m6 A-dependent manner [36]. Epigenetic changes in TME Recent epigenetics research has shifted its focus to the TME, notably in terms of immune system organization during tumor progression. The tumor microenvironment (TME) is made up of stromal cells, immune cells, extracellular matrix, and cytokines, and it creates a favorable and immune-suppressive environment for tumor cells. During tumor progression, epigenetic reprogramming is accompanied by changes in TME in both stromal compartments

and immune response, specifically the aberrant landscape of non-signaling RNAs. Exosomes, microvesicles, ectosomes, large oncosomes, and apoptotic vesicles are just a few of the extracellular vesicles generated by many cell types in TME, according to emerging evidence [37].

They contain DNA fragments mRNAs, and non-signaling RNAs and serve as essential communicative messengers between cells during premetastatic niche formation, EMT, and metastatic progression [38]. For example, exosomal miR-200b was higher in pancreatic ductal adenocarcinoma and was linked to a lower overall survival [39]. Exosomes from non-small cell lung malignant tumor cells boosted malignant tumor cell proliferation and metastasis by overexpressing miR-619-5p, which reduced rcan1.4 and increased malignant tumor cell proliferation and metastasis [40].

Furthermore, miRNAs that translocate to tumor cells can transmit oncogenic signals that shape favorable TME. To facilitate vascular permeability and angiogenesis, exosomal miR-25-3p was delivered to endothelial cells in the pre-metastatic niche of colon-rectal malignant tumor cells [41]. Malignant tumor DNA, mRNA, and miRNA in EVs likely retarded the phenotypic change of fibroblasts exposed to CRC-derived EVs [42]. MiR-409 and miR-154\*, which are generally suppressed after early embryogenesis, have been discovered to be upregulated in cancer associated fibroblasts [43].

Tumor progression requires an immune-suppressive environment in order for tumor cells to avoid immune surveillance and T-cell-mediated anti-malignant tumor death. The inhibitory immune checkpoint pathway is one technique for stopping the immune system. T-cells can be turned off by inducing certain checkpoint proteins, which is controlled by epigenetics in malignant tumors. Lower repressive histone marks and DNA methylation indicators are frequently detected at the all of checkpoint proteins [44, 45]. DMTASE1 inhibited cytotoxic T-cell homing to malignant tumor cells by lowering CXCL12 epigenetically in osteosarcomas [46].

Using epigenetics to treat malignant tumors malignant tumors are characterized by reversibility and sensitivity to extrinsic stimuli, and epigenetic modifications play a critical role in their formation. They're showing to be promising cancer therapeutic targets. For more than 40 years, epigenetic medications, or pharmaceuticals that target the epigenome, have been available. They have previously been investigated in clinical trials for the treatment of malignant tumors and have showed some promise. Cytosine analogues can bind to the DNA or RNA backbone and replace the C-5 of cytosine with N-5, causing methylation to be disrupted and DMTASE to be broken. Among them are 5-aza-cytidine, 5-aza-2'-deoxycytidine, zebularine, SGI-110, fazarabine, and others. Azacytidine and decitabine have been approved by the FDA as cytosine analogue inhibitors for the treatment of hematologic malignancies [47].

They are now often linked to a wide range of solid tumors decitabine is exclusively incorporated into DNA, whereas the majority of azacytidine incorporated into RNA. The activity of decitabine begins with DNA integration. The azacytosine-guanine dinucleotide that is produced captures dmtases with irreversible covalent connections, depleting them and removing DNA methylation marks on TSG [48]. This method also activates the DNA damage response, which results in cell cycle arrest, growth slowed, and death. Recent investigations have revealed that azacytidine can impede gene translation by changing tRNA-rRNA interactions and reducing deoxyribonucleotide conversion due to its ability to integrate RNA [49]. The anti-tumor capabilities of these two drugs have been determined in clinical studies at very low dosages due to the considerable toxicity induced by big doses. Zebularine, six-thioguanine, and four'-thio-two'-deoxycytidine are some more cytosine analogues that function with several ways than azacytidine and decitabine. ZEB incorporates a 2-(1H)-pyrimidine ring after DNA integration that promotes dmtase degradation by forming a covalent compound with dmtase at the six part of the pyrimidine ring [50, 51], the ZEB is not effective as azacytidine or decitabine due to the competitive action of cytidine deaminase, but it aids in preventing are methylation of the gene after other dmtase type one therapy and may reduce dmtaseis dosage [52, 53]. They are small molecules that bind to the catalytic site of dmtases or the CpG regions to prevent dmtases from binding to target sequences. Non-nucleotide analogue inhibitors are a type of non-nucleotide analogue inhibitor. Among them are hydralazine, aRG108, aMG98, and disulfiram. These epigenetic medications have a minor inhibitory effect on a variety of aggressive tumor cells when compared to cytosine analogue inhibitors. MG98 is an antisense oligonucleotide that targets the 3'UTR of dmtase1 and promotes demethylation [54].

Important suppression of DMTASE1 coding was identified in a phase one clinical investigation. Also, in a phase II clinical trial, no effect was seen in people with metastatic kidney cancer. Furthermore, HDACI-induced apoptosis is more vulnerable to malignant tumor cells. They are a good target for cancer therapy because of these properties. Based on their structure, HDACIs can be divided into four classes [55]. However, no impact was detected in persons with metastatic kidney cancer in a phase II clinical trial. By retriggering TSGs, HDAC inhibitors can restore the abnormal acetylation state of histones and non-histone proteins in malignant tumors. Furthermore, HDACI-induced apoptosis makes malignant tumor cells more susceptible. They're a good cancer treatment target because of these characteristics. HDACIs can be classified into four classes based on their structure: HDACIs have a hydroxamic acid

moiety that can bind to the zinc atom found in HDACs' catalytic sites, rendering them inactive. They have been demonstrated to be successful in the treatment of both hematologic and solid malignancies in numerous investigations. Three generic hydroxamic acid HDACIs were recently authorized by the FDA [56, 57].

HDACI that produces histone acetylation but it is nonselective. It has been demonstrated to increase the coding of p21 in bladder cancer and endometrial stromal sarcomas by inducing acetylated histone H3 and H4 [58, 59].

Glutathione reduces it within cells, releasing a zinc-binding FK2280 interacts more directly with this thiol-containing class one and two HDACs, resulting in reactive target genes [60]. Valproic acid is an aliphatic fatty acid HDACI that particularly targets HDACs in classes one and two. It was originally created to treat epilepsy, but it was later utilized to anti-tumor therapy due to its ability to inhibit the proliferation and induce the differentiation of malignant tumor cells. VPA can promote histone H3 and H4 acetylation as well as caspase demethylation of target and tumor-specific antigens [61]. VPA is a promising epi-drug because of its low toxicity, good tolerance, and stability. Phenylbutyrate, AR-42, and pivanex are other short-chain fatty acid HDACIs (AN-9) In the 1990s, Suzuki *et al.*, published the first benzamide derivative, which inhibited HDACs considerably [62]. The benzamide HDACIs entinostat and tacedinaline (CI-994) are particularly active. They have a  $\alpha$ -aminophenyl group that binds to the particular location of class one HDACs and inhibits the enzyme [63]. In the first and second phase clinical studies, it was also well tolerated by patients with lymphoid malignancies and solid tumors. CI-994, meantime, has undergone phase two clinical studies and could be used alone or in combination with other chemotherapeutic medicines to treat solid tumors patients [64]. Extra terminal inhibitors and bromodomain inhibitors (BETIs) BRD2, BRD3, BRD4, and BRDT are members of the BET family of proteins, which feature two N-terminal tandem bromodomains and a C-terminus. To promote transcriptional activity, they form a complex with HDACs and other proteins. Because of its ability to assemble on both hyper-acetylated agene allows and "super-enhancers" to allow RNA-pol type two mediated transcriptional initiation and elongation, BRD4 is the most well-studied BET implicated in transcriptional organization and malignant tumor growth [65]. Several oncogenes have been identified as BRD4 effectors [66]. BETIs work by disrupting the interaction between BETs and acetylated histones. Several BETIs, including the thienodiazepine JQ1, I-BET762 have shown promising clinical outcomes with low toxicity and high activity. JQ1 has been shown to preferentially disrupt BRD2/3/4 and acetylated histone iJQ1 causes down regulation of one of the target genes, c-Myc, in a variety of malignant tumors [67]. Furthermore, by blocking aBRD4-mediated androgen receptor recruitment and transactivation, it has a significant anti-tumor effect even in castration-resistant cells. JQ1 has been shown to boost cytotoxic T-cell response by raising PD-L1 coding in recent research [68, 69]. OTX015 administration may reduce CSC phenotype, making it an appealing therapy in the most aggressive PC [70].

Inhibitors of histone methyltransferases/demethylase PRC2's enzymatic activity is controlled by EZH2, a histone methyltransferase in several subtypes of malignant tumors [71]. EPZ-6438, GSK2816126, and aCPI-1205 are small molecule EZH2 inhibitors that have been studied in several studies and have shown antineoplastic effects in both hematologic and solid tumors [72].

FDA Orphan Drug designation for malignant was tumors as well as lymphoma and follicular lymphoma. Histone demethylase inhibitors, such as LSD1 and Jumonji inhibitors, are another type of epigenetic drug precursor. The impairment of H3K4 demethylation is the basis for LSD1 inhibitors' action [73, 74]. LSD1 inhibitors are divided to: first class is reversible polypeptide or monoamine oxidase inhibitor derivatives, second class is irreversible monoamine oxidase inhibitor derivatives, third class is rationally designed fusions of active molecules, and last class is the new compounds not previously known to block monoamine oxidase. Highly specific aLSD1 inhibitors, such as HCI-2509 and NCL-1, have been discovered to be reversible inhibitors with a strong ability to prevent H3K4 demethylation.

In castration-resistant PC cells, these LSD1 inhibitors can cause a rogenin-dependent growth stop with no apparent side effects in vitro and in vivo [75]. Such inhibitors have been created by targeting the a2-OG of the JmjC family, which is a crucial component for demethylation of methylated lysine [76]. They include hydroxamate derivatives, pyridine dicarboxylates, Noxalyl amino acid derivatives, and compounds that interfere with metal binding. ncRNAs miRNAs have increasingly been identified as biomarkers for a variety of malignant cancers, with aberrant levels being used to determine pathology and prognosis [77].

Many more miRNA-based medications, such as liposome-formulated and micro RNA type 34a, are in clinical studies and may be turned into FDA-approved drugs in the future. In the treatment of certain malignant cancers, they either act as miRNA mimics or as miRNA antagonists. For example, tumor suppressor miRNA-34a has been found as a tumor suppressor with various targets which coding is usually relatively low in diverse malignant tumors, such as PC, NSCLC, and ALL. MRX34 has been refined as a unique method for increasing miR-34a levels in malignant tumor therapy, and a phase II clinical trial has been begun [78]. In treatment trials, antagomiR-mediated suppression of particular miRNAs is highly suggested in terms of increased code of onco-miRNAs in various cancers. AntagomiR-

214 was found to reduce disease severity in CTCL in a recent study, providing another perspective on CTCL therapy [79]. Although aJQ1 could also down regulate miR-214, this suggests that miR-214 up regulation was slowed by abnormal histone acetylation [80]. Following that, numerous ncRNAs are discovered to be abnormally expressed in various cancers. Few ncRNAs have been identified as medicines for therapeutic therapy due to their complex organizations and intricate processes. However, because they are allegedly stable in body fluid, they can be used to determine the severity of specific malignant tumors. The presence of more ncRNA PCA3 in urine, for example, may indicate the severity of a prostate cancer [81]. There are still numerous barriers to overcome in terms of clinical application of ncRNAs; nonetheless, introducing such non-coding RNAs with effective delivery mechanisms is always a viable option in malignant tumor therapy. The synergistic activities of epigenetic medications explain the impact of numerous epigenetic pharmacological therapies. For example, inducing immunogenic cell death with SAHA and panobinostat co-therapy showed substantial anti-malignant tumor action in colon adenocarcinoma and leukemia. Chemotherapy has remained a standard treatment option for advanced malignant tumors that cannot be surgically removed [82, 83].

One reason is that some chemotherapy drugs can cause abnormal epigenetic modifications following treatment. Cisplatin therapy, for example, can cause hyper methylation of numerous genes in ovarian malignant tumors, resulting in acquired resistance [84]. activating of those epigenetic silenced genes, decitabine can reduce and even reverse cisplatin resistance, implying that combining epigenetic drugs with other chemotherapeutic agents can not only allow potent tumor progression suppression, but also are-sensitize tumor cells to radiotherapy and chemotherapy. HDAC3 and HDAC5 mRNAs were shown to be elevated in HCC investigations. Inhibitors such HDACs can enhance histone H3 and HSP90 acetylation and upregulated CDH1 [85, 86] the most promising inhibitors, which have been approved for specific cancer therapy. The lack of surface indicators for antigen presentation, reduced T-cell response, are-educated TME, and other factors, however, limit the therapeutic benefit of ICIs. Epigenetic reprogramming can help overcome some of these barriers. The efficacy of anti-PD-1 antibodies can be potentiated by utilizing DNMTase inhibitors, according to recent research. The major histocompatibility class one signaling genes were methylated and maintained in quiet code state in breast malignant tumors. Guadecitabine, a DNMTase type one, has been shown to improve MHC type one code and promote CD8+ T cell infiltration in TME [87]. These reactions may enhance subsequent responses to anti-PD-1 antibodies. Another example is decitabine's ability to make CD8+ T cells more sensitive to anti-PD-1 antibodies by inhibiting DNMTase3A-mediated methylation in fatigued T cells [88]. Belinostat treatment was found to suppress CTLA-4, resulting in M1-phenotypic tumor-associated macrophages and a decrease in splenic regulatory T cells [89]. Many other epigenetic medicines have been shown to boost anti-tumor immune response by improving T-cell persistence in many malignant cancers, including lung malignant tumor, TNBC, and lymphoma [90].

## REFERENCES

1. Waddington C. The evolution of an evolutionist. Edinburgh: Edinburgh Univ. Press; 1975.
2. Bird A. Perceptions of epigenetics. *Nature*. 2007; 447: 396.
3. Egger G, Liang G, Aparicio A, Jones PA. Epigenetics in human disease and prospects for epigenetic therapy. *Nature*. 2004; 429: 457.
4. Easwaran H, Tsai H-C, Baylin SB. Cancer epigenetics: tumor heterogeneity, plasticity of stem-like states, and drug resistance. *Mol Cell*. 2014; 54:716–27.
5. Lister R, Pelizzola M, Dowen RH, Hawkins RD, Hon G, Tonti-Filippini J, Nery JR, Lee L, Ye Z, Ngo Q-M. Human DNA methylomes at base resolution show widespread epigenomic differences. *Nature*. 2009; 462: 315.
6. Gopalakrishnan S, Van Emburgh BO, Robertson KD. DNA methylation in development and human disease. *Mut Res Fundam Mol Mech Mutagenesis*. 2008;647: 30–8.
7. Goyal R, Reinhardt R, Jeltsch A. Accuracy of DNA methylation pattern preservation by the Dnmt1 methyltransferase. *Nucleic Acids Res*. 2006;34: 1182–8.
8. Liang G, Chan MF, Tomigahara Y, Tsai YC, Gonzales FA, Li E, Laird PW, Jones PA. Cooperativity between DNA methyltransferases in the maintenance methylation of repetitive elements. *Mol Cell Biol*. 2002; 22: 480–91.
9. Scourzac L, Mouly E, Bernard OA. TET proteins and the control of cytosine demethylation in cancer. *Genome Med*. 2015; 7: 1–16.
10. Ke S, Alemu EA, Mertens C, Gantman EC, Fak JJ, Mele A, Haripal B, Zucker-Scharff I, Moore MJ, Park CY. A majority of m6A residues are in the last exons, allowing the potential for 3' UTR regulation. *Genes Dev*. 2015; 29: 2037–53.
11. Chen X-Y, Zhang J, Zhu J-S. The role of m6A RNA methylation in human cancer. *Mol Cancer*. 2019; 18: 103.
12. Meyer KD, Jaffrey SR. Rethinking m6A readers, writers, and erasers. *Annu Rev Cell Dev Biol*. 2017; 33: 319–42.
13. Müller S, Glaß M, Singh AK, Haase J, Bley N, Fuchs T, Lederer M, Dahl A, Huang H, Chen J. IGF2BP1 promotes SRF-dependent transcription in cancer in a m6A- and miRNA-dependent manner. *Nucleic Acids Res*. 2018; 47: 375–90.
14. Offermanns S, Rosenthal W. *Encyclopedia of molecular pharmacology*. Berlin, Heidelberg: Springer Science & Business Media; 2008.
15. Wiles ET, Selker EU. H3K27 methylation: a promiscuous repressive chromatin mark. *Curr Opin Genet Dev*. 2017; 43: 31–7.
16. Cavalli G. EZH2 Goes Solo. *Science*. 2012; 338: 1430–1.

17. Djebali S, Davis CA, Merkel A, Dobin A, Lassmann T, Mortazavi A, Tanzer A, Lagarde J, Lin W, Schlesinger F. Landscape of transcription in human cells. *Nature*. 2012; 489: 101–8.
18. Friedman RC, Farh KK-H, Burge CB, Bartel DP. Most mammalian mRNAs are conserved targets of microRNAs. *Genome Res*. 2009;19:92–105.
19. Jeck WR, Sharpless NE. Detecting and characterizing circular RNAs. *Nat Biotechnol*. 2014; 32: 453.
20. Wu P, Mo Y, Peng M, Tang T, Zhong Y, Deng X, Xiong F, Guo C, Wu X, Li Y. Emerging role of tumor-related functional peptides encoded by lncRNA and circRNA. *Mol Cancer*. 2020; 19: 22.
21. Vessoni AT, Filippi-Chiela EC, Lenz G, Batista LFZ. Tumor propagating cells: drivers of tumor plasticity, heterogeneity, and recurrence. *Oncogene*. 2020; 39: 2055-68.
22. Majumdar S, Buckles E, Estrada J, Koochekpour S. Aberrant DNA methylation and prostate cancer. *Curr Genomics*. 2011; 12: 486–505.
23. Sastry NG, Wan X, Huang T, Alvarez AA, Pangen RP, Song X, James CD, Horbinski CM, Brennan CW, Nakano I. LY6K promotes glioblastoma Tumorigenicity via CAV-1-mediated ERK1/2 signaling enhancement. *Neuro Oncol*. 2020.
24. He J, Zhou M, Li X, Gu S, Cao Y, Xing T, Chen W, Chu C, Gu F, Zhou J. SLC34A2 simultaneously promotes papillary thyroid carcinoma growth and invasion through distinct mechanisms. *Oncogene*. 2020; 39: 2658-75.
25. Xiao C, Wu G, Zhou Z, Zhang X, Wang Y, Song G, Ding E, Sun X, Zhong L, Li S. RBBP6, a RING finger-domain E3 ubiquitin ligase, induces epithelial–mesenchymal transition and promotes metastasis of colorectal cancer. *Cell Death Dis*. 2019; 10: 1–17.
26. Ding X, He M, Chan AW, Song QX, Sze SC, Chen H, Man MK, Man K, Chan SL, Lai PB. Genomic and Epigenomic Features of Primary and Recurrent Hepatocellular Carcinomas. *Gastroenterology*. 2019; 157: 1630–45 e1636.
27. Blanco E, González-Ramírez M, Alcaine-Colet A, Aranda S, Di Croce L. The bivalent genome: characterization, structure, and regulation. *Trends Genet*. 2019; 36: 118-31.
28. Zaidi SK, Fietze SE, Gordon JA, Heath JL, Messier T, Hong D, Boyd JR, Kang M, Imbalzano AN, Lian JB. Bivalent epigenetic control of oncofetal gene expression in cancer. *Mol Cell Biol*. 2017; 37: e00352–17.
29. Lhoumaud P, Badri S, Rodriguez-Hernaez J, Sakellaropoulos T, Sethia G, Kloetgen A, Cornwell M, Bhattacharyya S, Ay F, Bonneau R. NSD2 overexpression drives clustered chromatin and transcriptional changes in a subset of insulated domains. *Nat Commun*. 2019; 10: 1–18.
30. Rezaei T, Mansoori B, Hashemi ZS, Amini M, Rezaei S, Karami H, Mosafer J, Mokhtarzadeh A, Baradaran B. microRNA-181 serves as a dual-role regulator in the development of human cancers. *Free Radic Biol Med*. 2019.
31. Gregory PA, Bert AG, Paterson EL, Barry SC, Tsykin A, Farshid G, Vadas MA, Khew-Goodall Y, Goodall GJ. The miR-200 family and miR-205 regulate epithelial to mesenchymal transition by targeting ZEB1 and SIP1. *Nat Cell Biol*. 2008; 10: 593.
32. Guil S, Esteller M. RNA-RNA interactions in gene regulation: the coding and noncoding players. *Trends Biochem Sci*. 2015; 40: 248–56.
33. Han C, Fu Y, Zeng N, Yin J, Li Q. LncRNA FAM83H-AS1 promotes triplenegative breast cancer progression by regulating the miR-136-5p/ metadherin axis. *Aging*. 2020; 12. <https://doi.org/10.18632/aging.102832>.
34. He Q, Yan D, Dong W, Bi J, Huang L, Yang M, Huang J, Qin H, Lin T. circRNA circFUT8 upregulates Krüppel-like factor 10 to inhibit the metastasis of bladder Cancer via sponging miR-570-3p. *Mol Ther Oncolytics*. 2020; 16: 172–87.
35. Pan Y, Ma P, Liu Y, Li W, Shu Y. Multiple functions of m 6 a RNA methylation in cancer. *J Hematol Oncol*. 2018; 11: 48.
36. Alarcón CR, Lee H, Goodarzi H, Halberg N, Tavazoie SF. N 6- methyladenosine marks primary microRNAs for processing. *Nature*. 2015; 519: 482–5.
37. Jurj A, Zanoaga O, Braicu C, Lazar V, Tomuleasa C, Irimie A, BerindanNeagoe I. A comprehensive picture of extracellular vesicles and their contents. *Molecular Transfer to Cancer Cells*. *Cancers*. 2020; 12: 298.
38. Ni J, Bucci J, Malouf D, Knox M, Graham P, Li Y. Exosomes in Cancer Radioresistance. *Front Oncol*. 2019; 9: 869.
39. Reese M, Flammang I, Yang Z, Dhayat SA. Potential of Exosomal microRNA200b as liquid biopsy marker in pancreatic ductal adenocarcinoma. *Cancers*. 2020; 12: E197.
40. Kim, D. H., Park, S., Kim, H., Choi, Y. J., Kim, S. Y., Sung, K. J., ... & Rho, J. K. (2020). Tumor-derived exosomal miR-619-5p promotes tumor angiogenesis and metastasis through the inhibition of RCAN1. 4. *Cancer letters*, 475, 2-13.
41. Zeng Z, Li Y, Pan Y, Lan X, Song F, Sun J, Zhou K, Liu X, Ren X, Wang F. Cancer-derived exosomal miR-25-3p promotes pre-metastatic niche formation by inducing vascular permeability and angiogenesis. *Nat Commun*. 2018; 9: 5395.
42. Abdouh M, Floris M, Gao Z-H, Arena V, Arena M, Arena GO. Colorectal cancer-derived extracellular vesicles induce transformation of fibroblasts into colon carcinoma cells. *J Exp Clin Cancer Res*. 2019; 38: 257.
43. Josson S, Gururajan M, Sung S-Y, Hu P, Shao C, Zhau H, Liu C, Lichterman J, Duan P, Li Q. Stromal fibroblast-derived miR-409 promotes epithelial-tomesenchymal transition and prostate tumorigenesis. *Oncogene*. 2015; 34: 2690.
44. Zahavi DJ, Weiner LM. Targeting multiple receptors to increase checkpoint blockade efficacy. *Int J Mol Sci*. 2019; 20: 158.
45. Sasidharan Nair V, El Salhat H, Taha RZ, John A, Ali BR, Elkord E. DNA methylation and repressive H3K9 and H3K27 trimethylation in the promoter regions of PD-1, CTLA-4, TIM-3, LAG-3, TIGIT, and PD-L1 genes in human primary breast cancer. *Clin Epigenetics*. 2018; 10: 78.

46. Li, B., Wang, Z., Wu, H., Xue, M., Lin, P., Wang, S., ... & Ye, Z. (2018). Epigenetic Regulation of CXCL12 Plays a Critical Role in Mediating Tumor Progression and the Immune Response In Osteosarcoma OS Fate Determined by Epigenetic Regulation of CXCL12. *Cancer Research*, 78(14), 3938-3953.
47. Kantarjian H, Issa JPI, Rosenfeld CS, Bennett JM, Albitar M, DiPersio J, Klimek V, Slack J, De Castro C, Ravandi F. Decitabine improves patient outcomes in myelodysplastic syndromes: results of a phase III randomized study. *Cancer Interdiscip Int J Am Cancer Soc*. 2006; 106: 1794–803.
48. Stresmann C, Lyko F. Modes of action of the DNA methyltransferase inhibitors azacytidine and decitabine. *Int J Cancer*. 2008; 123: 8–13.
49. Schaefer M, Hagemann S, Hanna K, Lyko F. Azacytidine inhibits RNA methylation at DNMT2 target sites in human cancer cell lines. *Cancer Res*. 2009; 69: 8127–32.
50. Zhou L, Cheng X, Connolly B, Dickman M, Hurd P, Hornby D. Zebularine: a novel DNA methylation inhibitor that forms a covalent complex with DNA methyltransferases. *J Mol Biol*. 2002; 321: 591–9.
51. Holleran JL, Parise RA, Joseph E, Eiseman JL, Covey JM, Glaze ER, Lyubimov AV, Chen Y-F, D'Argenio DZ, Egorin MJ. Plasma pharmacokinetics, oral bioavailability, and interspecies scaling of the DNA methyltransferase inhibitor, zebularine. *Clin Cancer Res*. 2005; 11: 3862–8.
52. Tan W, Zhou W, Yu H-G, Luo H-S, Shen L. The DNA methyltransferase inhibitor zebularine induces mitochondria-mediated apoptosis in gastric cancer cells in vitro and in vivo. *Biochem Biophys Res Commun*. 2013; 430: 250–5.
53. Cheng JC, Weisenberger DJ, Gonzales FA, Liang G, Xu G-L, Hu Y-G, Marquez VE, Jones PA. Continuous zebularine treatment effectively sustains demethylation in human bladder cancer cells. *Mol Cell Biol*. 2004; 24: 1270–8.
54. Yang X., Lay F, Han H, Jones PA. Targeting DNA methylation for epigenetic therapy. *Trends Pharmacol Sci*. 2010; 31: 536–46.
55. Mann BS, Johnson JR, Cohen MH, Justice R, Pazdur R. FDA approval summary: vorinostat for treatment of advanced primary cutaneous T-cell lymphoma. *Oncologist*. 2007; 12: 1247–52.
56. McDermott J, Jimeno A. Belinostat for the treatment of peripheral T-cell lymphomas. *Drugs Today (Barcelona, Spain: 1998)*. 2014; 50: 337–45.
57. Richardson PG, Laubach JP, Lonial S, Moreau P, Yoon S-S, Hungria VT, Dimopoulos MA, Beksac M, Alsina M, San-Miguel JF. Panobinostat: a novel pan-deacetylase inhibitor for the treatment of relapsed or relapsed and refractory multiple myeloma. *Expert Rev Anticancer Ther*. 2015; 15: 737–48.
58. Hrzenjak A, Moinfar F, Kremser M-L, Strohmaier B, Petru E, Zatloukal K, Denk H. Histone deacetylase inhibitor vorinostat suppresses the growth of uterine sarcomas in vitro and in vivo. *Mol Cancer*. 2010; 9: 49.
59. Frye R, Myers M, Axelrod KC, Ness EA, Piekarczyk RL, Bates SE, Booher S. Romidepsin: a new drug for the treatment of cutaneous T-cell lymphoma. *Clin J Oncol Nurs*. 2012; 16: 195-204.
60. Xiao JJ, Byrd J, Marcucci G, Grever M, Chan KK. Identification of thiols and glutathione conjugates of depsipeptide FK228 (FR901228), a novel histone protein deacetylase inhibitor, in the blood. *Rapid Commun Mass Spectrom*. 2003; 17: 757–66.
61. Milutinovic S, D'Alessio AC, Detich N, Szyf M. Valproate induces widespread epigenetic reprogramming which involves demethylation of specific genes. *Carcinogenesis*. 2007; 28: 560–71.
62. Kato S, Morie T, Kon T, Yoshida N, Karasawa T, Matsumoto J. Novel benzamides as selective and potent gastrokinetic agents. 2. Synthesis and structure-activity relationships of 4-amino-5-chloro-2-ethoxy-N-[[4-(4-fluorobenzyl)-2-morpholinyl] methyl] benzamide citrate (AS-4370) and related compounds. *J Med Chem*. 1991; 34: 616–24.
63. Hess-Stumpff H, Bracker TU, Henderson D, Politz O. MS-275, a potent orally available inhibitor of histone deacetylases--the development of an anticancer agent. *Int J Biochem Cell Biol*. 2007; 39: 1388–405.
64. Witta SE, Jotte RM, Konduri K, Neubauer MA, Spira AI, Ruxer RL, VarellaGarcia M, Bunn PA Jr, Hirsch FR. Randomized phase II trial of erlotinib with and without entinostat in patients with advanced non-small-cell lung cancer who progressed on prior chemotherapy. *J Clin Oncol*. 2012; 30: 2248.
65. Donati B, Lorenzini E, Ciarrocchi A. BRD4 and Cancer: going beyond transcriptional regulation. *Mol Cancer*. 2018; 17: 164.
66. Sancisi V, Manzotti G, Gugnoni M, Rossi T, Gandolfi G, Gobbi G, Torricelli F, Catellani F, Faria do Valle I, Remondini D. RUNX2 expression in thyroid and breast cancer requires the cooperation of three non-redundant enhancers under the control of BRD4 and c-JUN. *Nucleic Acids Res*. 2017; 45: 11249–67.
67. Ott CJ, Kopp N, Bird L, Paranal RM, Qi J, Bowman T, Rodig SJ, Kung AL, Bradner JE, Weinstock DM. BET bromodomain inhibition targets both cMyc and IL7R in high-risk acute lymphoblastic leukemia. *Blood*. 2012; 120: 2843–52.
68. Zhu H, Bengsch F, Svoronos N, Rutkowski MR, Bitler BG, Allegranza MJ, Yokoyama Y, Kossenkov AV, Bradner JE, Conejo-Garcia JR. BET bromodomain inhibition promotes anti-tumor immunity by suppressing PD-L1 expression. *Cell Rep*. 2016; 16: 2829–37.
69. Vázquez R, Riveiro ME, Astorgues-Xerri L, Odore E, Rezai K, Erba E, Panini N, Rinaldi A, Kwee I, Beltrame L. The bromodomain inhibitor OTX015 (MK8628) exerts anti-tumor activity in triple-negative breast cancer models as single agent and in combination with everolimus. *Oncotarget*. 2017; 8: 7598.
70. Civenni G, Pedrani S, Allegrini S, Bruccoleri A, Albino D, Pinton S, GarciaEscudero R, Ouafik LH, Cvitkovic E, Carbone GM. Targeting prostate cancer stem cells (CSCs) with the novel BET bromodomain (BRD) protein inhibitor OTX015. Philadelphia: AACR; 2015.

71. Xu B, Konze KD, Jin J, Wang GG. Targeting EZH2 and PRC2 dependence as novel anticancer therapy. *Exp Hematol*. 2015; 43: 698–712.
72. Knutson SK, Warholic NM, Wigle TJ, Klaus CR, Allain CJ, Raimondi A, Scott MP, Chesworth R, Moyer MP, Copeland RA. Durable tumor regression in genetically altered malignant rhabdoid tumors by inhibition of methyltransferase EZH2. *Proc Natl Acad Sci*. 2013; 110: 7922–7.
73. Gulati N, Béguelin W, Giulino-Roth L. Enhancer of zeste homolog 2 (EZH2) inhibitors. *Leuk Lymphoma*. 2018; 59: 1574–85.
74. Forneris F, Binda C, Dall'Aglio A, Fraaije MW, Battaglioli E, Mattevi A. A highly specific mechanism of histone H3-K4 recognition by histone demethylase LSD1. *J Biol Chem*. 2006; 281: 35289–95.
75. Gupta S, Weston A, Bearrs J, Thode T, Neiss A, Soldi R, Sharma S. Reversible lysine-specific demethylase 1 antagonist HCl-2509 inhibits growth and decreases c-MYC in castration- and docetaxel-resistant prostate cancer cells. *Prostate Cancer Prostatic Dis*. 2016; 19: 349–57.
76. Yi T, Fang J, Erdjument-Bromage H, Warren ME, Borchers CH, Tempst P, Zhang Y. Histone demethylation by a family of JmjC domain-containing proteins. *Nature*. 2006; 439: 811.
77. Wood H. FDA approves patisiran to treat hereditary transthyretin amyloidosis. *Nature Reviews Neurology*. 2018; 14: 570.
78. Hanna J, Hossain GS, Kocerha J. The potential for microRNAs in clinical research. *Front Genet*. 2019; 10: 478.
79. Kohnken R, McNeil B, Wen J, McConnell K, Grinshpun L, Keiter A, Chen L, William B, Porcu P, Mishra A. Preclinical targeting of MicroRNA-214 in cutaneous T-cell lymphoma. *J Invest Dermatol*. 2019; 139: 1966-74.
80. Lee B, Mazar J, Aftab MN, Qi F, Shelley J, Li J-L, Govindarajan S, Valerio F, Rivera I, Thurn T. Long noncoding RNAs as putative biomarkers for prostate cancer detection. *J Mol Diagnostics*. 2014; 16: 615–26.
81. West AC, Mattarollo SR, Shortt J, Cluse LA, Christiansen AJ, Smyth MJ, Johnstone RW. An intact immune system is required for the anticancer activities of histone deacetylase inhibitors. *Cancer Res*. 2013; 73: 7265–76.
82. Welsbie DS, Xu J, Chen Y, Borsu L, Scher HI, Rosen N, Sawyers CL. Histone deacetylases are required for androgen receptor function in hormonesensitive and castrate-resistant prostate cancer. *Cancer Res*. 2009; 69: 958–66.
83. Harding T, Swanson J, Van Ness B. EZH2 inhibitors sensitize myeloma cell lines to panobinostat resulting in unique combinatorial transcriptomic changes. *Oncotarget*. 2018; 9: 21930.
84. Zeller C, Dai W, Steele NL, Siddiq A, Walley AJ, Wilhelm-Benartzi C, Rizzo S, Van Der Zee A, Plumb J, Brown R. Candidate DNA methylation drivers of acquired cisplatin resistance in ovarian cancer identified by methylome and expression profiling. *Oncogene*. 2012; 31: 4567.
85. Lachenmayer A, Toffanin S, Cabellos L, Alsinet C, Hoshida Y, Villanueva A, Minguez B, Tsai H-W, Ward SC, Thung S. Combination therapy for hepatocellular carcinoma: additive preclinical efficacy of the HDAC inhibitor panobinostat with sorafenib. *J Hepatol*. 2012; 56: 1343–50.
86. Bitzer M, Horger M, Giannini EG, Ganten TM, Wörns MA, Siveke JT, Dollinger MM, Gerken G, Scheulen ME, Wege H. Resminostat plus sorafenib as second-line therapy of advanced hepatocellular carcinoma—the SHELTER study. *J Hepatol*. 2016; 65: 280–8.
87. Luo N, Nixon MJ, Gonzalez-Ericsson PI, Sanchez V, Opalenik SR, Li H, Zahnow CA, Nickels ML, Liu F, Tantawy MN. DNA methyltransferase inhibition upregulates MHC-I to potentiate cytotoxic T lymphocyte responses in breast cancer. *Nat Commun*. 2018; 9: 248.
88. Ghoneim HE, Fan Y, Moustaki A, Abdelsamed HA, Dash P, Dogra P, Carter R, Awad W, Neale G, Thomas PG. De novo epigenetic programs inhibit PD-1 blockade-mediated T cell rejuvenation. *Cell*. 2017; 170: 142–57 e119.
89. Llopiz D, Ruiz M, Villanueva L, Iglesias T, Silva L, Egea J, Lasarte JJ, Pivette P, TrochonJoseph V, Vasseur B. Enhanced anti-tumor efficacy of checkpoint inhibitors in combination with the histone deacetylase inhibitor Belinostat in a murine hepatocellular carcinoma model. *Cancer Immunol Immunother*. 2019; 68: 379–93.
90. Sheng W, LaFleur MW, Nguyen TH, Chen S, Chakravarthy A, Conway JR, Li Y, Chen H, Yang H, Hsu P-H. LSD1 ablation stimulates anti-tumor immunity and enables checkpoint blockade. *Cell*. 2018; 174: 549–63 e519.